

Anti-Tumor Activity of *Phoenix dactylifera L.* Pit Extracts against *Hela* and *L₂₀B* cell lines

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Abstract

Date palm has antioxidant activity which is able to control cancer cells. This study was designed to assess the anti-tumor activity of aqueous and ethanolic extracts of *Phoenix dactylifera L.* pits extracts against (*Hela*) and (*L₂₀B*) cancer cell lines. HPLC technique detected that both extracts contained several phenolic compounds (Caffeic acid, Ferulic acid, *p*-coumaric acid, Chlorogenic acid, Sinapic acid, Genstic acid, *p*-hydroxy benzoic acid, Gallic acid and Protocatechuic acid). The percentage of growth inhibition (PGI) of four extracts concentrations (6.5, 12.5, 25 and 50 µg/ml) was in vitro assessed with using two cancer cell lines.

The results revealed that the four concentrations of the two plant extracts had anti-tumor activity in a concentration-dependent manner and the ethanolic extract gave best results of PGI and IC₅₀ than aqueous extract in both cell lines. As a conclusion in accordance to the results, the ethanolic extract of *Phoenix dactylifera L.* pits showed obvious anti-tumor activity in both cell lines. This activity appeared to be particularly significant in both treated cell lines.

Keywords: Date Palm pits, Anti-Tumor Activity, Human cell line, mice cell line.

Introduction

Date palm (*Phoenix dactylifera L.*) is considered as one of the most important source of food for humans in many regions¹¹. Its contain sugars reaching to 88% in some varieties⁷, also rich in vitamins and mineral salts and different chemical compounds such as Calcium (Ca), Cadmium (Cd), Zinc (Zn), Potassium (K), Saturated and Unsaturated fatty acids. The pits powder is also used in some traditional medicines and has been tested for human used for health purposes¹²

Cancer is the abnormal cellular growth that usually invades and destroys normal cells. As a result of imbalance in the body these cells are born and so various types of cancer treatments were used to correct the imbalance. In spite of many cancer researches, the nature of cancer does not understand exactly¹³. Thus, all over the world cancer kills about 3500 M people⁹. More than 270,000 people in USA die because of cancer⁴. In Iraq, breast cancer considered to

be the most dominant type of all cancers representing around 43% with a mortality rate of 23%⁸.

Cancer therapy depends on surgery radiotherapy, chemotherapy, or combinations of them. New and /or complementary treatments methods are needed as a result of significant death rate in response to cancer and because of the serious side effects of radiation therapy and chemotherapy. Because of the need for effective chemotherapy and in line with recent discoveries in cell biology to cure cancer without serious side effects, recent researchers are doing their best¹⁹.

Considered as a major treatment, chemotherapy is used for the control of advanced stages of cancer cells yet shows serious side effects on normal cells and tissues^{19,27}.

Since the beginning of civilizations, the plant has been used to treat many diseases for both humans and animals. Without causing toxicity, they may maintain the vitality and health of people and also cure diseases including cancer⁶. About 50% of all modern drugs in medicinal use are from natural products which have the ability to treat tumor cells²⁴.

Phenolic compounds in plants have a major role in control cancer cells¹⁷, because some synthetic antioxidants have the ability to enhance development of cancer in mice and the increased interested of peoples in natural foods products. Plant phenolic compounds and other natural antioxidants are extremely desirable².

Material and Methods

Collection of the Plant: The date palm pits were freshly collected from Iraqi local strains Barhi and Khastawi of date palm in year 2017.

Preparation of aqueous and ethanolic extracts of *Phoenix dactylifera L.* pits for anti-tumor activity: Pits of *Phoenix dactylifera L.* were ground into fine powder using grinding machine³.

Aqueous extract: About 50 g of the powdered pits was mixed with 250 ml double distilled water. Then mixture was left in a shaker incubator at 37°C for 24 hrs, then filtered by a filter paper (Whatmann no. 1). Using rotary evaporator, the filtrate was concentrated at 40°C until dryness and the extract residue was weighed and kept until used²⁸.

Ethanolic extract: About 50 g of the powdered pit was extracted with 250 ml of 70% ethanol in flask and put on

magnetic stirrer for 72 hrs. at room temperature, then filtered through filter paper (Whatmann no.1). Using a rotary evaporator, the solution was evaporated to dryness at 40°C and the extract residue was weighed and kept until used¹⁶.

Detection of some phenolic compounds of date palm aqueous pits extract:

- Estimation of phenols date palm pit by HPLC: The main compound was separated on FLC (Fast Liquid Chromatography) column under the optimum condition.
- Column: phenomenex C-18, 3µm particle size (100x 4.6 mm I.D) column.
- Mobile phase: Linear gradient of solvent A 0.1 % trifluoro acetic acid (TFA acid) in deionized water: solvent B was acetonitrile gradient program from 0% B to 100 % B for 10 minutes.
- Flow rate 1.3 ml/min.
- Detection: UV at 280 nm.
- Calculation:
Concentration of sample µg/ml = (sample area / standard area) x conc. of standard x dilution Factor.
- Extraction: 10 mg of the extraction phenolic compounds dissolved in 10 ml methanol were exposed to ultra – sonication (Branson sonifier, USA) at 60 % duty cycles for 25 min. at 25 °C and centrifugation (7500 rpm for 15 min.), then to remove pigments prior to evaporation under vacuum (Buchi Rotary evaporator Re Type) the clear supernatant of sample was subjected to charcoal treatment. Re-suspend the dried samples in 1.0 ml methanol by vortex, the mixture was passed through 2.5 µm disposable filter and stored at 4°C for further analysis, then according to the optimum separation condition, 20 µl of the sample was injected into HPLC system^{5,20}.

Assessment of Cytotoxic Activity (*in vitro*): The *in vitro* anti-tumor activity of agrimony aqueous and ethanol extracts was estimated at the Biotechnology Research Center (Al-Nahrain University). In this study, the cytotoxic activity of the two pits extracts was determined against two tumor cell lines (Human cell line (HELA) and (L₂₀B) mice cell line)¹⁵. The percentage of growth inhibition was calculated according to an equation presented by Phuangsab et al.²³

$$\text{Growth inhibition (\%)} = \left(\frac{\text{Control Absorbance} - \text{Treated Absorbance}}{\text{Control Absorbance}} \right) \times 100$$

Growth Inhibition Assessment in HeLa and L₂₀B Cell Lines: The laboratory assessment of growth inhibition was carried out according to a method that was adopted by Abdul-Majeed¹. The method included the following steps:

1. The cells (Hela and L₂₀B) were supplemented as monolayer attached cells in Falcon culture flasks (75 cm) containing RPMI-1640 medium. The cells were washed with PBS and then 1 ml of trypsin – versine solution was added with a gentle shaking until the cells were detached from the

flask surface. Such manipulation was carried out with aid of phase contrast inverted microscope. Then, the contents of the flask were removed to another flask and incubated at 37°C for 15 minutes (sub-culture).

2. The cells were counted and their number was adjusted to 1 x 10⁶ cell/ml. At the same time, viability was assessed using a dye-exclusion test (trypan blue stain) and it was always greater than 96%.
3. The cells were seeded in the wells of 96-well tissue culture plate, which was carried out by pipetting 200 µl of the cell suspension into each well in a cell number of 10⁴ cells/ well and the plate was incubated overnight at 37°C.
4. The day after, the wells were examined to inspect the cell growth, then four concentrations 6.5, 12.5, 25 and 50 µg/ml were added to each well in a triplicate manner.
5. After the end of each incubation period, 20 µl of MTT solution was added to each well. The plate was re-incubated at 37°C for 2-4 hrs.
6. After incubation, DMSO was added to dissolve the MTT crystals.
7. The wells were read under the ELISA reader at wave length of 570 nm and the absorbance was recorded.

Statistical Methods: The values of the resulted parameters were given in terms of mean and the differences between means were assessed by analysis of variance test, using the computer program SPSS (using T-test). The difference was considered significant when the probability value was equal or less than 0.05.

Results and Discussion

Result estimated by HPLC methods showed that palm pits extracts contain a significant amount of some important phenolic compounds with higher concentrations for Caffeic acid and Ferulic acid respectively (table 2). Both tested strains showed similar results in aqueous and ethanolic extracts.

Both tested cell lines show similar results of PGI in aqueous and ethanolic extracts. Results in table 3 and figure 1 showed that ethanolic extract has the best cytotoxic activity with highest PGI at the higher concentration used (83.8 % at 50 µg/ml) than that obtained by aqueous extract (70 % at 50 µg/ml).

Nevertheless, the calculated IC₅₀ values showed no significant differences between both different extracts against HeLa cells and slightly lower value for the aqueous extract.

Cytotoxic effects of date palm pits extracts (aqueous and ethanol) on L₂₀B cell line: The results in table 4 and figure

2 showed that ethanolic extract has the best anti-tumor activity with highest PGI at the higher concentration used (85.7 % at 50 µg/ml) than that obtained by aqueous extract (77.1 % at 50 µg/ml). Nevertheless, the calculated IC₅₀ value was significantly different and had the lower value for the aqueous extract.

Plants are sources of different compounds. They contain and can produce a variety of chemical substances that have different biological actions with a special reference to their medicinal importance^{6,21}. Therefore, they are employed by herbalists of different cultures, anciently and recently to provide remedy to peoples of their sicknesses¹⁰.

Since cancer is a major cause of death, every day tumor treated research is conducted worldwide. These studies often involve the properties and the effects of biologically active compounds on tumor cells and they often originate from plants²⁵. There is an urgent need to examine reliable and inexhaustible sources of natural compounds. In addition, it is important to understand the mechanisms of anti-tumor activity for future application in cancer treatment¹⁴.



Figure 1: Hela cell line after treated them by date palm seeds extract (24 hour).

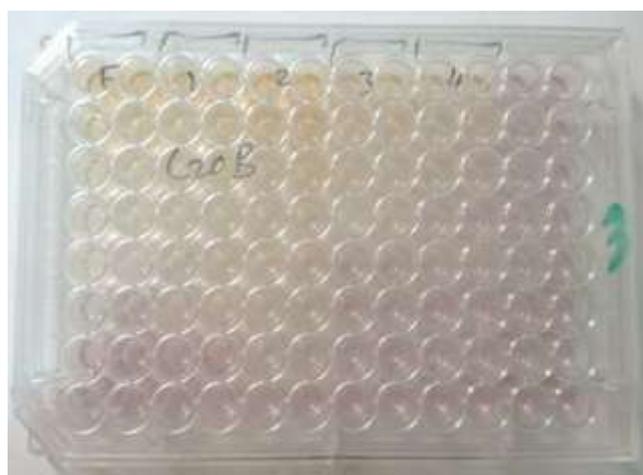


Figure 2: L₂₀B cell line after treated them by date palm seeds extract (24 hour).

Table 1
Sequences of the eluted material of the standard (Concentration 25 µg / ml).

Seq.	Subjects	Retention time / minute	Area µvolt
1	Gallic acid	1.28	34652
2	Genstic acid	2.12	70529
3	Cafeic acid	3.04	95867
4	Chlorogenic acid	5.53	81401
5	p-coumaric acid	6.35	106120
6	Ferulic acid	7.43	118751
7	p-hydroxy benzoic acid	8.52	98922
8	Sinapic acid	9.24	84973
9	Protocatechuic acid	10.39	119875

Table 2
Estimation of some active compounds (phenolic compounds) in Palm pit aqueous extracts

Phenolic compounds	Concentration µg / ml
Caffeic acid	276.33
Ferulic acid	235.12
Chlorogenic acid	164.26
p-coumaric acid	145.16
Sinapic acid	95.09
Genstic acid	91.42
p-hydroxy benzoic acid	91.11
Gallic acid	78.57
Protocatechuic acid	19.44

Table 3
Cytotoxic effects of date palm seed (aqueous and methanol) extract against Hela cell line.

Extract Concentration (µg/ml)	Growth Inhibition (%)	
	Aqueous Extract	Ethanol Extract
6.5	40.3	48.8
12.5	43.8	50.5
25	66.8	58.3
50	70.0	83.8
IC ₅₀	7.38	7.80

This research investigated the anti-tumor activity of the ethanolic and aqueous extract (in a concentration of 6.5 -12.5 -25 -50 µg/mL) on HELA and L₂₀B cell lines. A dose-dependent inhibition was seen in treated cell line. For functional antagonist and competitive binding assays, the

most common measurement of the dose-response curve is the IC₅₀. The half maximal inhibitory concentration (IC₅₀) is an estimation of the effect of a substance in inhibiting a specific biological or biochemical function.

Table 4
Cytotoxic effects of date palm seed extracts
(aqueous and ethanol) on L₂₀B cell line.

Extract Concentration (µg/ml)	Percentage of Growth Inhibition	
	Aqueous extract	Ethanol extract
6.5	26.0	38.7
12.5	34.6	48.6
25	35.7	70.0
50	77.1	85.7
IC ₅₀	5.3	6.2

The cell proliferation was significantly lower when compared to untreated control cells after 24 h of treatment but lower concentrations had lower anti-tumor activity. Maximum inhibition of proliferation was seen at the highest concentration (50 µg/mL). The anti-tumor activity may be attributed to the presence of compounds that can inhibit cell cancer found naturally in this plant especially the most active antioxidative compounds of date palm pits are phenolic compounds pointed in detection (Caffeic acid, Ferulic acid, Chlorogenic acid, Sinapic acid, p-coumaric acid, Genstic acid, p-hydroxy benzoic acid, Gallic acid, Protocatechuic acid) in different concentrations^{11,22}. Many researches showed that phenolic compounds have a significant role in cancer treatments^{18,26,29}.

Conclusion

Many active compounds were detected in the aqueous and ethanolic extracts of date palm pits including Caffeic acid, Sinapic acid, Ferulic acid, Chlorogenic acid, p-coumaric acid, Genstic acid, p-hydroxy benzoic acid, Gallic acid, Protocatechuic acid. Anti-tumor activities of both date palm pits extracts were *in vitro* detected with different concentrations but the ethanolic extract was more efficient than aqueous extract.

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References

- Abdul-Majeed M.R., Induction and Characterization of SU99 Plasmacytoma Cell Line and Its Effects on Mice Immune Response, Ph.D. Thesis, College of Science, AL-Nahrain University, Iraq (2000)
- Aberoumand A. and Deokule S.S., Comparison of phenolic compounds of some edible plants of Iran and India, *Pak J Nutr.*, **7**, 582–585 (2004)

- Abu-Shanab B., Adwan G., Abu-Safiya D., Jarrar N. and Adwan K., Antibacterial activities of some plant extracts utilized in popular medicine in Palestine, *Turk. J. Biol.*, **28(2-4)**, 99-102 (2004)
- Ahmedin J., Rebecca S., Elizabeth W., Taylor M., Jiaquan X. and Michael J., Cancer Statistics, American Cancer Society, CA, *Cancer J. Clin.*, **57**, 43-66 (2007)
- Al-Farsi M.A. and Lee C.Y., Optimization of phenolics and dietary fibre extraction from date seeds, *Food Chem.*, **108**, 977–985 (2008)
- Al-Saffar Ali Z., Al-Shanon Ahmed F., Al-Brazanchi Shymaa L., Sabry Fatimah A., Hassan Firas and Hadi Noora A., Phytochemical Analysis, Antioxidant and Cytotoxic Potentials of *Pelargonium graveolens* Extract in Human Breast Adenocarcinoma (MCF-7) Cell Line, *Asian Journal of Biochemistry*, **12(1)**, 16-26 (2017)
- Al-Shahib W. and Marshall R.J., Fatty acid content of the seeds from 14 varieties of date palm *Phoenix dactylifera* L., *Int J Food Sci Tech.*, **38**, 70912 (2003)
- Alwan N.A.S., Breast Cancer among Iraqi Women, Preliminary Findings from a Regional Comparative Breast Cancer Research Project, *J. Global Oncol.*, **2(5)**, 255-258 (2016)
- American Cancer Society, A biotechnology company dedicated to cancer treatment, viewed on 25 January, www.cancervax.com/info/index.htm (2006)
- Anderson F.J., An illustrated history of herbals, Columbia University Press, New York (1997)
- Besbes S., Blecker C., Deroanne C., Drira N.E. and Attia H., Date seeds: chemical composition and characteristic profiles of the lipid fraction, *Food Chem.*, **84**, 577-584 (2004)
- Booij G., Piombo J.M., Risterucci M., Coupe D. and Ferry M., Study of the chemical composition of dates at various stages of maturity for varietals characterization of various of date palm cultivars (*Phoenix dactylifera* L.), *Fruits*, **47**, 667-677 (1992)
- Estrogen and cancer website, www.womenshealth.com (2006)
- Filip Grbović, Stanković Milan S., Milena Ćurčić, Nataša Đorđević, Dragana Šeklić, Marina Topuzović and Snežana Marković, *In Vitro* Cytotoxic Activity of *Origanum vulgare* L. on HCT-116 and MDA-MB-231 Cell Lines, *Plants*, **2**, 371-378 (2013)
- Freshney R.I., Culture of animal cells, A manual of basic technique and specialized applications, 6th ed., John Wiley and Sons, Inc. USA (2010)
- Harborne J.B., Phytochemical Methods, Chapman & Hall, London (1998)
- Khanavi M., Saghari Z., Mohammadirad A., Khademi R., Hadjiakhoondi A. and Abdollahi M., Comparison of antioxidant activity and total phenols of some date varieties, *DARU Journal of Pharmaceutical Sciences*, **17**, 104-107 (2009)

18. Livia Brenelli de Paiva, Rosana Goldbeck, Wanderley Dantas dos Santos and Fabio Marcio Squina, Ferulic acid and derivatives: molecules with potential application in the pharmaceutical field, *Brazilian Journal of Pharmaceutical Sciences*, **49(3)**, 395–411 (2013)
19. Madhuri S. and Pandey G., Some anticancer medicinal plants of foreign origin, *Current Science*, **96(6)**, 779-783 (2009)
20. Obouayeba Abba Pacôme, Djyh Nazaire Bernard, Diabate Sékou, Djaman Allico Joseph, N'guessan Jean David, Kone Mongomaké and Kouakou Tanoh Hilaire, Phytochemical and Antioxidant of Roselle (*Hibiscus sabdariffa* L.) Petal Extracts, *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, **4(5)**, 1454 (2014)
21. Pandey G. and Madhuri S., Medicinal plants: better remedy for neoplasm, *Indian Drugs*, **43**, 869–874 (2006)
22. Pengxuan Zhang, Yuping Tang, Nian-Guang Li, Yue Zhu and Jin-Ao Duan, Bioactivity and Chemical Synthesis of Caffeic Acid Phenethyl Ester and Its Derivatives, *Molecules*, **19**, 16458-16476 (2014)
23. Phuangsab A., Lorence R.M., Reichard K.W., Peebles M.E. and Walters R.J., Newcastle disease virus therapy of human tumor xenografts: in vitro effects of local or systematic administration, *Cancer Lett.*, **172(1)**, 27-36 (2001)
24. Rosangkima G. and Prasad S.B., Antitumour activity of some plants from Meghalaya and Mizoram against murine ascites Dalton's lymphoma, *Indian J. Exp. Biol.*, **42**, 981–988 (2004)
25. Salih Shahlaa M., Alobaidi Khalid H. and Alobaidi Zinah F., Cytotoxic Effect of *Rosmarinus officinalis* L. Leaf Extracts on Tumor Cell Line, *Journal of Al-Nahrain University / Science*, **18(4)**, 98-102 (2015)
26. Sayed Gad El Molla, Amira Abdel Motaal, Hala El Hefnawy and Ahlam El Fishawy, Cytotoxic activity of phenolic constituents from *Echinochloa crus-galli* against four human cancer cell lines, *Revista Brasileira de Farmacognosia*, **26(1)**, 62–67 (2016)
27. Somkumar A.P., Studies on anticancer effects of *Ocimum sanctum* and *Withania somnifera* on experimentally induced cancer in mice, Ph.D. Thesis, JNKVV, Jabalpur, India (2003)
28. Swanston F.S.K., Day C., Baileg C.J. and Flatt P.R., Technique of plant analysis, Chapman Hall, London. Traditional plant treatments for diabetes. Studies in normal and streptozotocin diabetic mice, *Diabetologia*, **33**, 462-464 (1990)
29. Zhang P., Tang Y., Li N.G., Zhu Y. and Duan J.A., Bioactivity and Chemical Synthesis of Caffeic Acid Phenethyl Ester and Its Derivatives, *Molecules*, **19**, 16458-16476 (2014).