Computational based Characterization of Heat shock protein Hsp27 from Humans and Canines

Saleem Afnan*, Singh Satparkash and Sunil Kumar B.V. Division of Animal Biotechnology, Faculty of Veterinary Sciences and Animal Husbandry, Sher-e-Kashmir University of Agricultural Sciences and Technology-Kashmir, INDIA *afnankhan1082@gmail.com

Abstract

Heat shock protein functions as molecular chaperones with an imperative role in diverse cellular processes including protein folding, actin organization and stress resistance. Heat shock protein 27 (Hsp27), a member of the small Hsp (sHsp) has been linked to tumor progression as well as a therapeutic target for cancer treatment in humans as well as canines. A detailed in silico computational investigation of the Hsp27 protein from humans and canines was analyzed with respect to its structural, functional and phylogenetic properties. Although less extensively characterized in canines, attempts were made to compare the human protein with that of canines using the computational tools available to help the researchers get acquainted with the protein structure. Hsps are the most evolutionary conserved class of proteins.

Nevertheless, the Hsp27 proteins of humans and canines are clearly related. Though divergent in sequence, Hsp27 proteins are conserved in their structural properties. Hsp27 protein also appears to share other functional properties as well. In this study, a flexible, unstable, hydrophillic protein with an average molecular weight of 27KDa was reported for humans as well as canines. Specifically, the phosophorylation of protein supports the development of anti-Hsp27 agents for treatment of cancer. Moreover, this theoretical in silico analysis of Hsp27 protein will be substantial for cancer research and health care.

Keywords: Heat shock protein, hsp27, protein, cancer, analysis, canine.

Introduction

Heat shock proteins or Hsps are a group of proteins that are produced in response to heat. This response mechanism is the most highly conserved genetic system known, existing in all the known organisms from archaebacteria to eubacteria, from animals to plants, even though the response might vary in different organisms. Hsps were first initially studied in response to heat shock at elevated temperatures³¹ but are now acknowledged to express during stresses like cold exposure²⁴, tissue remodeling or wound healing²² and UV light⁴. Hsps play a vital role in normal cell functioning as well as functioning as intra cellular chaperones by ensuring correct folding of the proteins⁹.

Based on their molecular weights, Hsp proteins are classified as Hsp70, Hsp90 where 70 stands for size of the protein in kilodaltons²³. Ubiquitin is a small 8-Kda protein when induced by heat, it also produces the heat shock response³⁰. Small heat shock protein (sHSP) is a 80 amino-acid alpha crystallin conserved protein binding domain²¹. Proteins are ubiquitously the most abundant proteins induced by heat shock as well as in normal development. Besides their role in thermotolerance, they also play an important part in cell proliferation, cancer vaccines and therapeutics, drug resistance and agriculture which make them proteins of special clinical interest.

Heat shock protein 27 (Hsp27) is a member of the small Hsp (sHsp) family among α -crystallin, Hsp20 and others and is also known as heat shock protein beta-1 (HSPB1)⁵. The common functions of sHSPs include thermotolerance, actin organization, inhibition of apoptosis, cell differentiation and regulation of cell development⁴⁰. The sHps are reported as highly pleiotropic molecules both in terms of cell biology reports and molecular function. High expression levels of Hsp 27 are found in various cancers⁷ including the sera of breast cancer patients indicative of metastasis and resistance to chemotherapy³³. Hsp27 levels are triggered in stress by transcriptional activation and post-translational modification (phosphorylation) subsequential of the p38 MAPK stress kinase pathway³.

Hsp27 has also been shown to be a mediator in cancer development and progression being a vital player of angiogensis³⁹. A recent study reported the elevated serum Hsp27 in dogs with mammary tumors as compared to healthy dogs irrespective of the mammary tumor histotypes suggesting expoitation of Hsp27 as neoplastic signature of canine mammary tumors¹. Hsp27 also appears to serve a significant role in various muscle/neurodegenerative diseases³⁴. Notably, heritable mutations in Hsp27 central Pro residue have been reported to cause the hereditary motor neuropathy Charcot-Marie-Tooth disease¹⁰.

The goal of this manuscript is to analyze the structural and functional properties of the Hsp27 protein in humans and canines. Although less extensively characterized in canines, attempts were made to compare the human protein with that of canines using the computational tools available to help the researchers get acquainted with the protein structure.

Material and Methods

Sequence retrieval and Multiple sequence alignment (MSA): The protein sequences of the human and the canine heat shock protein Hsp27 were retrieved from the NCBI protein database. Basic Local Alignment Search Tool (BLASTp) was used to get the similar sequences of other organisms (cattle, buffalo, goat, pig). Accession numbers of the following proteins are presented in table I. The sequences once retrieved in FASTA format were aligned using Clustal omega online tool using default parameters.

Table I
Accession numbers of the Hsp27 protein

Species name	Accession number			
Homo Sapiens (Human)	CAA38016.1			
Canis lupus (Dog)	AXQ88113.1			
Bos Taurus (Cattle)	NP_001020740			
Bubalus bubalus (Buffalo)	AIU47315.1			
Capra hircus (Goat)	AFK93550.1			
Sus scrofa (Pig)	AAV54182.1			

Primary structural analysis of Hsp27 protein: Primary structural analysis of the Hsp27 protein of humans and canines was determined using ProtParam tool from Expert Protein Analysis System (ExPASy) which is the proteomic server of Swiss Institute of Bioinformatics (SIB) (https://www.expasy.org/)¹². The biophysical and biochemical properties include molecular weight (Mw), number of positive and negative residues, isoelectric point(pI), extinction coefficients (EC-quantitative study of protein-protein and protein-ligand interactions)¹³, instability index (II-stability of proteins)¹⁵, aliphatic index (AI-relative volume of protein occupied by aliphatic side chains)¹⁶, grand average hydropathicity (GRAVY-sum of all hydropathicity values of all amino acids divided by number of residues in a sequence)¹⁷ and half life¹⁴.

Secondary and tertiary structure prediction of Hsp27 protein: The amino acid sequence of the human and canine Hsp27 was subjected to secondary protein structure prediction using **PSIPRED** 4.0server (http://bioinf.cs.ucl.ac.uk/psipred/)¹⁷. The tertiary structure prediction of the Hsp27 protein was modelled using ab initio approach with online available tool Swiss model software (https://swissmodel.expasy.org/)⁴². The model thus obtained, was further validated by ramachandran's plot (http://mordred.bioc.cam.ac.uk/~rapper/rampage.php)

Functional analysis: Functional characterization of the human and canine Hsp27 for identifying protein families, functional sites and domains was done using Inter-ProScan (https://www.ebi.ac.uk/interpro/)¹⁸. ProtComp 9.0 identified the sub-cellular localization of proteins (ProtComp - Version 9). Motifs in the Hsp27 protein sequences were predicted by Psite online tool which is a protein domain database for functional annotation and description³⁶. Potential phosphorylation sites of the protein sequences were

predicted using NetPhos3.1² (http://www.cbs.dtu.dk/ services/NetPhosK) server that is provided by Centre for Biological Sequence Analysis, Technical University of Denmark (CBS DTU).

The server predicts serine, threonine or tyrosine phosphorylation sites in eukaryotic proteins using ensembles of neural networks. The SS bonding of the cysteine residues in the Hsp27 protein was predicted using CYC_REC tool (CYS_REC). Location of signal peptide cleavage sites was predicted using Signal P-4.1²⁷.

Phylogenetic analysis: Phylogenetic analysis helps to understand the evolutionary relationship among the diverse species. The phylogenetic tree was constructed using unweighted pair group method (UPGMA) of the Molecular Evolutionary Genetics Analysis version 6.0 (MEGA)¹⁹. The consistency of the inferred phylogenetic tree was checked by bootstrap analysis of 1000 replications. The gaps and missing data were eliminated.

Protein-protein interaction study: STRINGv10.0 web server (http://string-db.org) was used to predict the interaction of human and canine Hsp27 protein with their other closely allied proteins respectively. A critical assessment of the protein-protein interaction network based on the direct and indirect associations for both the proteins was generated³⁸.

Ethical approval: Since the study does not involve any animal study, animal ethics committee approval was not required. The study utilizes the available online computational tools and the online gene sequences.

Results

Sequence retrieval and Multiple sequence alignment (MSA): % similarity of the human and canine Hsp27 protein with other species was determined. Human hsp27 was found to be 88, 87, 86, 89 and 88 percent identical to dog, cattle, buffalo, goat and pig respectively. Similarly, canine hsp27 was also 88, 90, 90, 89 and 91 percent identical to human, cattle, buffalo, goat and pig respectively. Human protein had the highest percent identity with goat while the canine protein resembled with pig. Multiple sequence alignment using Clustal omega software suggests variability among different species (Figure I).

A remarkable feature of sHsps to note was that it showed greater homology within organisms than between organisms. A report of soybean small hsp family revealed 90% amino-acid identity with each other whereas 20% amino-acid identity with the proteins of D. melanogaster, C. elegans and X. laevis was reported²⁵.

Primary structural analysis of Hsp27 protein: Table II shows the amino acid composition of the human and canine Hsp27 protein as evaluated by the ProtParam online server. Human Hsp27 has 205 amino acids while the canine Hsp27

has 206 amino acids. Proline and serine are the most abundant amino acids in humans followed by alanine. In canines, proline was the most abundant amino acid in canines followed by alanine and serine. Both the proteins have a homologus and highly conserved amino acid sequence with similar structural features.

Human	MTERRVPFSLLRGPSWDPFRDWYP-HSRLFDQAFGLPRLPEEWSQWLGGSSWPGYVRPLP	59
Canine	MTERRVPFSLLRSPSWDPFRDWYPAHSRLFDQAFGLPRLPEEWAQWFGHSGWPGYVRPIP	60
Pig	MTERRVPFSLLRSPSWDPFRDWYPAHSRLFDQAFGLPRLPEEWSQWLSHSGWPGYVRPLP	60
Cattle	MAERRVPFSLLRGPSWDPFRDWYPAHSRLFDQAFGLPRLPEEWSQWLSHSGWPGYVRALP	60
Buffalo	MAERRVPFSLLRGPSWDPFRDWYPAHSRLFDQAFGLPRLPEEWSQWLSHSGWPGYVRALP	60
Goat	MAERRVPFSLLRGPSWDPFRDWYPAHSRLFDQAFGLPRLPEEWSQRLSHSGWPGYVRPLP	60
	*;***********;*************************	
Human	PAAIESPA-VAAPAYSRALSRQLSSGVSEIRHTADRWRVSLDVNHFAPDELTVKTKDGVV	118
Canine	P-AVEGPAAAAAPAYSRALSRQLSSGVSEIRQTADRWRVSLDVNHFAPEELTVKTKDGVV	119
Pig	PPAIEGPAAVAAPAYSRLLSRQLSSGVSEIQQTADRWRVSLDVNHFAPEELTVKTKDGVV	120
Cattle	AAAIEGPAYNRALSRQLSSGVSEIQQTADRWRVSLDVNHFAPEELTVKTKDGVV	114
Buffalo	AAAIEGPAYNRALSRQLSSGVSEIQQTADRWRVSLDVNHFAPEELTVKTKDGVV	114
Goat	AAAIEGPAYSRALSRQLSSGVSEIQQTADRWRVSLDVNHFAPEELTVKTKDGVV	114
	:.* **.* ***********:::**************	
Human	EITGKHEERQDEHGYISRCFTRKYTLPPGVDPTQVSSSLSPEGTLTVEAPMPKLATQSNE	178
Canine	EITGKHEERQDEHGYISRCFTRKYTLPPGVDPTLVSSSLSPEGTLTVEAPMPKPATQSAE	179
Pig	EITGKHEERQDEHGFISRCFTRKYTLPPGVDPTQVSSSLSPEGTLSVEAPLPKPATQSAE	180
Cattle	EITGKHEERQDEHGYISRCFTRKYTLPPGVDPTLVSSSLSPEGTLTVEAPLPKSATQSAE	174
Buffalo	EITGKHEERQDEHGYISRCFTRKYTLPPGVDPTLVSSSLSPEGTLTVEAPLPKSATQSAE	174
Goat	EITGKHEERQDEHGYISRCFTRKYTLPPGVDPTQVSSSLSPEGTLTVEAPLPKSATQSAE	174

Human	ITIPVTFESRAQLGGPEAAKSDETAAK 205	
Canine	ITIPVTFEARAQIGGPEAGKSEQSGAK 206	
Pig	ITIPVTFEARAQLGGTEAGKSEKPGTK 207	
Cattle	ITIPVTFQARAQLGGPEAGKSEQPETSKDP 204	
Buffalo	ITIPVTFQARAQLGGPEAGKSEQPENK 201	
Goat	ITIPVTFQA 183	

Figure I: Multiple sequence alignment of Hsp27 protein

Amino acid composition of Hsp27						
	Human	Hsp27	Canine	e Hsp27		
Amino acid	d No. of % of a		No. of	% of amino		
	amino acid	acid	amino acid	acid		
Ala (A)	17	8.3%	20	9.7%		
Arg (R)	16	7.8%	16	7.8%		
Asn (N)	2	1.0%	1	0.5%		
Asp(D)	10	4.9%	8	3.9%		
Cys (C)	1	0.5%	1	0.5%		
Gln (Q)	7	3.4%	8	3.9%		
Glu (E)	16	7.8%	17	8.3%		
Gly (G)	13	6.3%	15	7.3%		
His (H)	5	2.4%	5	2.4%		
Ile (I)	6	2.9%	7	3.4%		
Leu (L)	16	7.8%	13	6.3%		
Lys (K)	7	3.4%	7	3.4%		
Met (M)	2	1.0%	2	1.0%		
Phe (F)	7	3.4%	8	3.9%		
Pro (P)	21	10.2%	22	10.7%		
Ser (S)	21	10.2%	19	9.2%		
Thr (T)	14	6.8%	13	6.3%		
Trp (W)	6	2.9%	6	2.9%		
Tyr (Y)	5	2.4%	5	2.4%		
Val (V)	13	6.3%	13	6.3%		

Table II Amino acid composition of Hsp27

Table III shows the different physicochemical properties of the human and canine Hsp27 protein. Isoelectric point (pI) of a molecule is the pH at which it carries no net charge and is electrically neutral. The computed pI will help in choosing a buffer for protein purification and crystallization. pI of human Hsp27 protein was computed to be 5.98 signifying its acidic nature while the pI for canine Hsp27 was 6.23 suggestive of its mild acidic nature as compared to the human hsp27. As the value of instability index for both the Hsp27 protein was higher than 40, it signifies its unstable nature⁴¹.

Aliphatic index stands for the relative volume of protein occupied by its aliphatic side chains (Alanine, isoleucine, leucine and valine). Aliphatic index values are directly propotional to the structural stability of a protein¹⁶. Both the proteins have a higher aliphatic index which suggests that the proteins are thermostable. The grand average of hydropathy (GRAVY) value indicates the solubility of proteins wherein the hydrophobic and hydrophilic properties of each amino acid chain are considered thoroughly. A positive value indicates a hydrophobic protein while a negative value indicates hydrophilic protein²⁰.

A negative GRAVY score for both the Hsp27 proteins indicate its hydrophilic character and a better interaction of protein and water. The estimated half life was around 30 hrs. Positively charged residues are calculated by the total number of arginine (Arg) and lysine (Lys) while the negatively charged residues are calculated by the total number of aspartic acid (Asp) and glutamic acid (Glu). These are supportive in resolving the topology of protein²⁶. Though divergent in sequence, the Hsp27 proteins of humans and canines are conserved in their physicochemical properties. Both the proteins are flexible with an average molecular weight of 27Kda, unstable and similar hydropathy profiles. **Secondary and tertiary structure prediction:** The secondary structure prediction of the amino acid sequence of human Hsp27 and the canine Hsp27 was determined elaborately using the PSIPRED online server. Upon comparative structure analysis, it was observed that both the human and canine Hsp27 protein structure had dominant coil structural content followed by strand and then helix. 57% of total amino acids contributed to coils, 26% to strand and 15% to helix in human Hsp27 protein (Figure IIa) whereas, 59% of total amino acids contributed to coils, 24% to strand and 16% to helix in canine Hsp27 protein (Figure IIb).

The dominance of coil structural content in the protein might be attributed to the presence of proline, as being the most abundant amino acid found in both humans as well as canine amino acid sequence. Proline has an exceptional property of creating kinks in the polypeptide chains, thus disrupting the secondary structure and consequential of coiling.

The tertiary structures of the Hsp27 proteins were predicted through the SWISS-MODEL. A template sequence with a significant similarity with the query sequence is required to predict the three dimensional structure of the protein. In our study, the selected template sequence was 6dv5.1, a crystal structure of Heat shock protein beta-1. The sequence identity of the template sequence with the query sequence was 100% for human model whileas it was 88.29% for canine model.

The oligo state of both the predicted protein models was homo-24-mer. Based on QMEAN and Z score, a good quality model for human (Figure IIIa) and canine was selected (Figure IIIb). Analysis of Ramachandran plot revealed 73.72% of the amino acids were in favoured and 9.61% were outliers for human Hsp27 model (Figure IVa). 73.69% of the amino acids were in favoured and 9.23% were outliers for canine Hsp27 model (Figure IVb). Similar *in silico* evaluation of human small heat shock protein Hsp27 by homology modeling has been reported¹¹ whereas the canine protein has been poorly characterized.

Physicochemical properties of lysyl oxidase protein					
S.N.	Biophysical and biochemical	Human Hsp27	Canine Hsp27		
	Properties	protein	protein		
1.	No of amino acids	205	206		
2.	Molecular weight	22782.52	22765.54		
3.	Isoelectric point	5.98	6.23		
4.	Negatively charged residues	26	25		
	(Asp + Glu)				
5.	Positively charged residues (Arg + Lys)	23	23		
6.	Extinction coefficients	40450	40450		
7.	Abs 0.1%	1.775	1.777		
8.	Instability index:	62.82	64.47		
		(Unstable)	(Unstable)		
9.	Aliphatic index:	68.54	65.87		
10.	(GRAVY)	-0.567	-0.541		
11.	Half life	30 hrs	30hrs		

Table III Physicochemical properties of lysyl oxidase protein







Figure IIb: Secondary structure prediction of canine Hsp27 protein using PSIPRED Coil dominates the secondary structure elements followed by strand

Functional analysis: Both the mammalian proteins appear to share the functional properties as well. InterproScan predicted protein's family membership with small heat shock protein Hsp20 and alpha crystallin for both proteins. ProtComp 9.0 revealed that the sub-cellular localization of the protein in humans as well as canines was possibly multilocated – cytoplasm and nucleus.

Hsp27 has the property of being induced during different stages in development suggestive of its significant role in cell differentiation. Using the Psite software, it was found that the human protein sequence had somewhat similar motifs when compared with the canine protein except for ATP/GTP-binding site motif A (P-loop) found only in canine sequence at 193-200 amino acid residue.

Protein kinase C phosphorylation in both the mammalian proteins protects against a non apoptotic cell death, thus supporting the development of anti-Hsp27 agents for treatment of cancer. Motif regions are summarized in table IVa and IVb. Serine (21), threonine(14) and tyrosine (5) are predicted as potential phosphorylation sites in human protein while serine (19), threonine(13) and tyrosine (5) are predicted as potential phosphorylation sites in canine

protein. Phosphorylation is closely associated with protein activity and indicative of protein function regulation.

Hsp27 phosphorylation has been reported to resist TNF- α induced apoptosis and increase in the formation of oligomers²⁹. A single cysteine residue was found in both the protein sequence at position 137 in humans and position 138 in canines using the CYC_REC tool and is probably not SSbounded. Cysteine residues play a vital role in protein's thermostability. The SignalP 5.0 server predicts the incidence of signal peptides and the position of their cleavage sites. The likelihood of the signal peptide was around 0.0019 for human protein and 0.0021 for canine protein.

Phylogenetic analysis: The phylogenetic tree constructed using MEGA software showed that the human Hsp27 was phylogenetically more similar to goat and cattle sequence while the canine Hsp27 was more similar to buffalo and pig sequence (Figure V). Human Hsp27 is forming an independent clad together with cattle and goat whilst it clusters away from the canine Hsp27 sequence which is forming another independent clad with buffalo and pig sequence. To infer the evolutionary history of sHSPs, a small eukaryotic sHSP was isolated from the mycobacterium M. $leprae^{43}$ suggesting that the small HSP family might have existed for over a billion years.

Protein-protein interaction study: Proteins generally function by interaction forming protein complexes. This network of protein illuminates the important clues as to the functioning of novel proteins. Based on various network parameters like gene fusion, text mining, co-expression, co-occurence, neighborhood and databases, a protein interaction network with 10 potential interacting protein associates was depicted for human (figure VIa) and canine

Hsp27 protein (figure VIb). For human Hsp27, the closest interacting protein was found MAP kinase-activated protein kinase 2 (MAPKAPK2) and death domain-associated protein 6 (DAXX) while the distant interacting protein was MAP kinase-activated protein kinase 3 (MAPKAPK3) and estrogen receptor. The closest interacting protein for canine Hsp27 was mitogen-activated protein kinase-activated protein kinase 2 (MAPKAPK2) while the distant interacting protein for canine Hsp27 was mitogen-activated protein kinase-activated protein kinase 2 (MAPKAPK2) while the distant interacting protein kinase 2 (MAPKAPK2) while the distant interacting protein was polyadenylate-binding protein (PABPC1). Potential interacting protein associates of HSP27 protein are listed in figure VII a and b.



Figure IIIa: Tertiary structure prediction of human Hsp27 protein



Figure IIIb: Tertiary structure prediction of canine Hsp27 protein



Figure IVa: Structure Validation by Ramachnadran Plot (Human Hsp27 protein)



Figure IVb: Structure Validation by Ramachnadran Plot (Canine Hsp27 protein)

p38 MAP kinase when stimulated by different agents or mitogens or inflammatory cytokines like IL-1 β and TNF α , leads to the activation of MAPKAP kinases 2 and 3 phosphorylating the mammalian sHSPs³². Phosphorylation is the key mode of activation of Hsp27 which facilitates the chaperone process under cellular stress³². Phosphorylated Hsp27 block the apoptotic pathways by their interaction with other pro-apoptotic proteins such as DAXX and ASK1⁶.

Conclusion

Heat shock proteins are a group of proteins which not just regulate the normal physiology but are also important mediators in diseases including cardiovascular, wound healing and cancer. A long standing assumption about the heat shock response is their protective action against the toxic effects of heat and other stresses. These are the most evolutionary conserved class of molecules playing an imperative role in cellular homeostasis. However, the most baffling of them would be the small Hsps which are not just abundant but are universally disseminated proteins for normal development as well as the for the heat shock response. Hsp27 has been implicated in tumor invasiveness and appears to be a promising therapeutic target for cancer in humans as well as canines.



Figure V: Phylogenetic tree showing evolutionary relationship among different organisms using MEGA 6.0 software (Boot-straps: 1000 replicates)



Figure VIa: Protein interaction map of human Hsp27 protein



Figure VIb: Protein interaction map of canine Hsp27 protein

e Hares	Heat shock protein beta 1, timal heat shock protein which functions as a malecular chapterine probably maintaining denotized proteins in a folding: comparised static. Plays a cale in stress resolution and actio organization. Through its malecular chapterine activity may regulate instruments biological processes including the phosphorylation and the warral transport of neuroficament proteins (205 as).		1	durer .	17	
Predicted Fund	tional Partners:	1188	8	8	33	3
C MAINAING	ABAP kinase activated protein kinase 2; Stress activated servicitlescoles protein Annual modered to cylokite production, int.					0.945
e baxx	Death domain associated protein & Transcription compressor known to represe transcriptional potential of several survey).					6.991
C MAPK14	Mitopen activated protein Kinase T-E. Serine/Howmine Kinase which acts as an essential component of the KKP kinase sty-					0.990
e HSPAE	Heat should cognite 71 kBn protein; Molecular straperone implicated in a write variety of cellular processes, including prote.			٠	٠	2.984
CYDS	Optichrome is Electron carrier protein. The selilized flams of the optichrome is heree program accept an electron from the			٠	٠	0.984
MARKAINS	MAP know activated protein know 5. Turner appressor serine threamine protein know encoded in mTORC1 appulling a.		٠	٠		0.984
CDCSL	Call distance cycle 3-like protein (INA binding protein involved in cell cycle control. May act as a transcription activator. Co.		•			0.960
· HEIFATA	Heat about 70 kDs protein 13, Minimular stagement implicated in a wide variety of cellular processes, including protection .		-	٠		0.979
MAPKAPK3	MAP knows activated protein knows 3. Stress activated permit Preserve protein Knows incohed in cytokines, production, e.			٠	٠	0.974
@ E903	Extragon receptor, Nuclear hormone receptor. The startist hormones and their receptors are involved in the regulation of eu-					0.957

Figure VIIa: Screenshot from STRING server of predicting interacting proteins with the query sequence

Your input.						
e HSPBT	Heat abook protein betw 1. Breat tear shock protein which functions as a molecular chapterine probably mentalising deviational proteins in a full-leg-competent state. Plays a role or above reasonant and actin organization. Through the molecular chapterine activity may replace numerous biological processes including the phospharylation and the assess mapped of anomhisment proteins (206 as).		etterite	dates -		
Predicted Fund	tional Partners:	2888	8	ā,	22	ā.
В МАРКАРКІ	Mitogen activated protein kinase activated protein kinase 2 (307 as)			٠		0.984
# MAPKAPKS	Althogen activated protein kinase-activated protein kinase 5 (472 x4)			٠	٠	0.968
C MAPKAPKS	Althogen-activated protein Atrawe-activated protein kinase 3 (000 au)			٠		0.963
e HSPAD	Uncharacterized protein: Beilanga to the heat abook protein 70 family (648 as)			٠		0.948
COC607182	Uncharacterized protein; Belongs to the heat aback protein 70 Aeroly (0-K au)		۰.			0.048
en Histophy	Heterogeneous nuclear ritionackeprotein D (AU-rich element RNA binding protein 1, 37kDa) (355 au)			٠		0.930
C EIFAGR	Exampletic translation settlation factor 4 gamma, 1 (1421 as)					0.925
@ U8/C	Uncharacterized protein (2R1 as)					0.923
1000	(Incharacterized protein (229 au)			٠	+	0.923
# PARFICE	Polyavheredane-bourding postneirs: Bouls the polycid) tail of milliple (1988 and					0.004

Figure VIIb: Screenshot from STRING server of predicting interacting proteins with the query sequence

 Table IVa

 Motif regions present in the human Hsp27 protein

 No. of sites
 Amino acid residues

Motif information	No. of sites	Amino acid residues
cAMP- and cGMP-dependent protein kinase	1	140-143
phosphorylation site		
Protein kinase C phosphorylation site	4	2-4, 110-112,121-123, 139-141
Tyrosine kinase phosphorylation site	1	127-133
N-myristoylation site	2	147-152, 192-197
Microbodies C-terminal targeting signal	4	26-28, 74-76, 102-104, 187-189

 Table IVb

 Motif regions present in the canine Hsp27 protein

Motif information	No. of sites	Amino acid residues			
cAMP- and cGMP-dependent protein kinase	1	141-144			
phosphorylation site					
Protein kinase C phosphorylation site	4	2-4, 110-112,122-124, 140-142			
Tyrosine kinase phosphorylation site	1	128-134			
N-myristoylation site	2	148-153, 193-198			
Microbodies C-terminal targeting signal	4	27-29, 75-77, 103-105, 188-190			
ATP/GTP-binding site motif A (P-loop)	1	193-200			

Thus, understanding the structural and functional properties of the protein will further facilitate a better understanding of mechanism of enzyme action. A comparative analysis of the human and canine Hsp27 protein signifies that the protein is more conserved in its structural properties. A flexible, unstable and a hydrophilic protein with a molecular weight of 27KDa was found in both the species. Further, the homology in their functional property would be sufficient to suggest a common function. The predicted secondary and

tertiary structures will aid in shedding light on the biological functions of the protein. The study of Hsp27 protein can be further explored and utilized for beneficial therapeutic and diagnostic purposes aiding medical as well as animal experts.

References

1. Birdi R. et al, Circulating level of heat shock protein 27 is elevated in dogs with mammary tumors, *3 Biotech*, **9**, 229 (**2019**)

2. Blom N., Gammeltoft S. and Brunak S., Sequence- and structure-based prediction of eukaryotic protein phosphorylation sites, *J. Mol. Biol.*, **294**, 1351-1362 (**1999**)

3. Brunet Simioni M., De Thonel A., Hammann A., Joly A.L., Bossis G., Fourmaux E., Bouchot A., Landry J., Piechaczyk M. and Garrido C., Heat shock protein 27 is involved in SUMO-2/3 modification of heat shock factor 1 and thereby modulates the transcription factor activity, *Oncogene*, **28**, 3332–3344 (**2009**)

4. Cao Y., Ohwatari N., Matsumoto T., Kosaka M., Ohtsuru A. and Yamashita S., TGF-beta1 mediates 70-kDa heat shock protein induction due to ultraviolet irradiation in human skin fibroblasts, *Pflügers Archiv.*, **438**, 239–44 (**1999**)

5. Carper S.W., Rocheleau T.A. and Storm F.K., cDNA sequence of a human heat shock protein HSP27, *Nuc. Acid. Res.*, **18**, 6457 (**1990**)

6. Charette S.J., Lavoie J.N., Lambert H. and Landry J., Inhibition of Daxx-mediated apoptosis by heat shock protein 27, *Mol Cell Biol.*, **20**, 7602–7612 (**2000**)

7. Ciocca D.R. and Calderwood S.K., Heat shock proteins in cancer: diagnostic, prognostic, predictive and treatment implications, *Cell Stress Chap.*, **10**, 86–103 (**2005**)

8. CYS_REC: The Program for Predicting SS-bonding States of Cysteines and disulphide bridgesin Protein Sequences, http://www.softberry.com/berry.phtml?topic=cys_rec&group=pro grams&subgroup=prop

9. De Maio A., Heat shock proteins: facts, thoughts and dreams, *Shock*, **11**, 1–12 (**1999**)

10. Evgrafov O.V. et al, Mutant small heat-shock protein 27 causes axonal Charcot-Marie-Tooth disease and distal hereditary motor neuropathy, *Nat. Genet.*, **36**, 602–6 (**2004**)

11. Fossa P. and Cichero E., *In silico* evaluation of human small heat shock protein HSP27: Homology modeling, mutation analyses and docking studies, *Bioorg. Med. Chem.*, **23**, 3215-3220 (**2015**)

12. Gasteiger E., Hoogland C., Gattiker A., Duvaud S., Wilkins M.R., Appel R.D. and Bairoch A., Protein Identification and Analysis Tools on the ExPASy Server, The Proteomics Protocols Handbook, Humana Press, 571-607 (**2005**)

13. Gill S.C. and Hippel P.H.V., Calculation of Protein Extinction Coefficient from Amino Acid Sequence Data, *Analytical Biochem.*, **182**, 319-326 (**1989**)

14. Gonda D.K., Bachmair A., Wunning I., Tobias J.W., Lane W.S. and Varshavsky A., A Universality and structure of the N-end rule, *J. Biol. Chem.*, **264**, 16700–16712 (**1989**)

15. Guruprasad K., Reddy B.V.B. and Pandit M.W., Correlation between stability of a protein and its dipeptide composition: a novel approach for predicting in vivo stability of a protein from its primary sequence, *Protein Engineering*, **4**, 55-161 (**1990**)

16. Ikai A., Thermostability and aliphatic index of globular proteins, *J. Biochem.*, **88**, 1895-1898 (**1980**)

17. Jones D.T., Protein secondary structure prediction based on position-specific scoring matrices, *J. Mol. Biol.*, **292**, 195-202 (**1999**)

18. Jones P., Binns D. and Chang H.Y., InterProScan 5: genomescale protein function classification, *Bioinformatics*, **30**, 1236-40 (**2014**)

19. Kumar S., Stecher G., Li M., Knyaz C. and Tamura K., MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms, *Mol Biol and Evol.*, **35**, 1547-9 (**2018**)

20. Kyte J. and Doolittle R.F., A simple method for displaying the hydropathic character of a protein, *J Mol Biol.*, **157**, 105-132 (**1982**)

21. Lahvic J.L., Ji Y., Marin P., Zuflacht J.P., Springel M.W., Wosen J.E., Davis L., Hutson L.D., Amack J.D. and Marvin M.J., Small heat shock proteins are necessary for heart migration and laterality determination in zebrafish, *Developmental Biology*, **384**, 166–80 (**2013**)

22. Laplante A.F., Moulin V., Auger F.A., Landry J., Li H., Morrow G., Tanguay R.M. and Germain L., Expression of heat shock proteins in mouse skin during wound healing, *The Journal* of *Histochemistry and Cytochemistry*, **46**, 1291–301 (**1998**)

23. Li Z. and Srivastava P., Heat-shock proteins, Current Protocols in Immunology, Appendix 1, Appendix 1T (**2004**)

24. Matz J.M., Blake M.J., Tatelman H.M., Lavoi K.P. and Holbrook N.J., Characterization and regulation of cold-induced heat shock protein expression in mouse brown adipose tissue, *The American Journal of Physiology*, **269**, R38–47 (**1995**)

25. Nagao R.T., Kimpel J.A., Vierling E. and Key J.L., The heat shock response: a comparative analysis, In Oxford Surveys of Plant Molecular and Cell Biology, ed., Miflin B.J., Oxford, Oxford Univ. Press, 384 (**1986**)

26. Nakashima H. and Nishikawa K., Discrimination of intracellular and extracellular proteins using amino acid composition and residue-pair frequencies, *J Mol Biol.*, **238**, 54-61 (**1994**)

27. Peterson T.N., Brunak S., Heijne G. and Nielsen H., SignalP 4.0: discriminating signal peptides from transmembrane regions, *Nat Methods*, **8**, 785-786 (**2011**)

28. ProtComp - Version 9, Program for Identification of subcellular localization of Eukaryotic proteins, Animal/Fungi, http://www.softberry.com/berry.phtml?topic=protcompan&group =programs&subgroup=proloc (2016)

29. Qi Z., Shen L., Zhou H., Jiang Y., Lan L., Luo L. and Yin Z., Phosphorylation of heat shock protein 27 antagonizes TNF- α induced HeLa cell apoptosis via regulating TAK1 ubiquitination and activation of p38 and ERK signaling, *Cellular Signalling*, **26**, 1616–25 (**2014**)

30. Raboy B., Sharon G., Parag H.A., Shochat Y. and Kulka R.G., Effect of stress on protein degradation: role of the ubiquitin system, *Acta Biologica Hungarica*, **42**, 3–20 (**1991**)

31. Ritossa F., A new puffing pattern induced by temperature shock and DNP in drosophila, *Experimental*, **18**, 571–573 (**1962**)

32. Rogalla T., Ehrnsperger M., Preville X., Kotlyarov A., Lutsch G., Ducasse C., Paul C., Wieske M., Arrigo A.P., Buchner J. and Gaestel M., Regulation of Hsp27 oligomerization, chaperone function and protective activity against oxidative stress/tumor necrosis factor alpha by phosphorylation, *J. Biol. Chem*, **274**, 18947–56 (**1999**)

33. Rui Z., Jian-Guo J., Yuan-Peng T., Hai P. and Bing-Gen R., Use of serological proteomic methods to find biomarkers associated with breast cancer, *Proteomics*, **3**, 433–9 (**2003**)

34. Sarto C., Binz P.A. and Mocarelli P., Heat shock proteins in human cancer, *Electrophoresis*, **21**, 1218–26 (**2000**)

35. Sievers F., Wilm A., Dineen D., Gibson T.J., Karplus K., Li W., Lopez R., McWilliam H., Remmert M., Söding J., Thompson J.D. and Higgins D.G., Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega, *Mol Syst Biol.*, **7**, 539 (**2011**)

36. Solovyev V.V. and Kolchanov N.A., Search for functional sites using consensus, In Computer analysis of Genetic macromolecules, World Scientific, 16-21 (**1994**)

37. Sun X., Ou Z., Xie M., Kang R., Fan Y., Niu X., Wang H., Cao L. and Tang D., HSPB1 as a novel regulator of ferroptotic cancer cell death, *Oncogene*, **34**, 5617–25 (**2015**)

38. Szklarczyk D., Morris J.H., Cook H., Kuhn M. and Wyder S., STRINGv10.0: protein-protein interaction networks, integrated over the tree of life, *Nucleic Acids Res.*, **43**, 447-452 (**2015**)

39. Thuringer D. et al, Extracellular HSP27 mediates angiogenesis through Toll-like receptor 3, *FASEB J.*, **27**, 4169–4183 (**2013**)

40. Van Montfort R., Slingsby C. and Vierling E., Structure and function of the small heat shock protein/alpha-crystallin family of molecular chaperones, *Advances in Protein Chemistry*, **59**, 105–56 (2001)

41. Verma A., Kumar V.S. and Gaur S., Computational based functional analysis of *Bacillus* phytases, *Computational Biology* and Chemistry, **60**, 53-58 (**2016**)

42. Waterhouse A., Bertoni M., Bienert S., Studer G., Tauriello G., Gumienny R., Heer F.T., de Beer T.A.P., Rempfer C., Bordoli L., Lepore R. and Schwede T., SWISS-MODEL: homology modelling of protein structures and complexes, *Nucleic Acids Res*, **46**, 296-303 (**2018**)

43. Young D., Lathigra R. and Mehlert A., Stress-induced proteins as antigens in infectious diseases, UCLA Symposium on Molecular & Cellular Biology, Stress-Induced Proteins, ed., Seramisco J.R., Lindquist S.L. and Pardue M.L., New York, Liss, 294 (**1988**).

(Received 01st November 2021, revised 31st July 2022, 25th August 2022)