# Review Paper: A review of the antioxidant activity of Apium Graveolens

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## Abstract

Plants are an important source of natural active products that differ depending on the chemical components they contain. Since extracts and phytochemicals isolated from plants show biological activity in vitro and in vivo, today plants are used as alternative treatment sources.

Apium graveolens (celery) has powerful antioxidant properties to remove free radicals due to compounds such as coumarin, alkaloids, steroids, phenols, essential oils, sesquiterpene alcohols, caffeic acid, pcoumaric acid, ferulic acid, apigenin, luteolin, tannin, saponin and kaempferol. Celery with different compounds and different concentrations has various healing effects. The aim of this study was to review the antioxidant activity of celery.

Keywords: Apium graveolens L, celery, antioxidant activity.

### Introduction

The use of medicinal plants to treat illness has been common since ancient times. Many studies have shown the positive effects of various herbs and different parts of medicinal plants on cancer, infectious diseases, diabetes, atherosclerosis.<sup>30,37,39</sup> Phenolic and alkaloid compounds in plants and their effects such as antioxidant effects have been investigated in many studies such as cancer,<sup>2,18,41</sup> diabetes,<sup>16,23</sup> liver disorders,<sup>30</sup> coronary heart diseases etc.<sup>24,25</sup> Today herbal drugs are used as an alternative to chemical drugs due to their low side effects.

Celery (*Apium graveolens* L) is a plant from the apiaceae family and is one of the annual or perennial plants that grow throughout Europe, Africa and Asia<sup>15</sup>. Celery seeds are used as a condiment in the flavoring of food products possessing a characteristic aroma and pungent taste. There are a number of phthalide derivatives that give the celery essential oil a characteristic odor.<sup>19</sup>

Celery (*Apium graveolens*) is a medicinal plant in traditional medicine with numerous health benefits. Celery can prevent arthritis, rheumatism, gout, urinary tract inflammation and specifically rheumatoid arthritis with mental depression.<sup>5</sup> Celery, because of compounds such as caffeic acid, p-coumaric acid, ferulic acid, apigenin, luteolin, tannin, saponin and kaempferol, has powerful antioxidant characteristics to remove free radicals. Antioxidants with radical scavenging capacity are thought to have a potential protective effect against free radical damage. These

biomolecules inhibit oxidative reactions that prevent the formation of coronary and vascular diseases and tumors.<sup>19,26</sup>

This oxidative damage is the result of free radical action on, for instance, lipids or DNA. However, the commonly used synthetic antioxidants such as butylated hydroxyanisole (BHA) and butylated hydroxyl toluene (BHT), are limited by law because of their toxic effects and carcinogenicity.<sup>28</sup> The elimination of synthetic antioxidants in food applications has provided further impetus to explore the source of natural antioxidants. The objective of the present review is to highlight the antioxidant effects of *Apium graveolens*.

### **Phytochemical Constituents**

The preliminary phytochemical analysis revealed the presence of carbohydrates, flavonoids, alkaloids, steroids and glycosides in the methanolic extract of seeds of Apium graveolens<sup>24</sup>. Seeds included flavonoids, volatile oils, coumarins and furanocoumarins. Coumarins contained celerin, bergapten, apiumoside, apiumetin, apigravrin, osthenol, isopimpinellin, isoimperatorin, celereoside and 5 and 8-hydroxy methoxypsoralen. Flavonoid included apiin, apigenin, isoquercitrin.<sup>5,20</sup> The phenolic concentration in different extracts(methanol, ethanol, water) varied significantly. Among the methanol extract of the seeds. Apium graveolens methanolic extract had the highest phenolic concentration  $(73.1 \pm 1.23 \text{ mg GAE}/100 \text{g}).^4$ Volatile oils included limonene (60%) and selenine (10-15%) and various sesquiterpene alcohols (1-3%), e.g.  $\alpha$ eudesmol and ß-eudesmol, santalol.

Also celery includes linoleic, myristic, myristoleic, oleic, palmitic, palmitoleic, petroselinic and stearic acid.<sup>5</sup> The main chemical constituents present in each part of the plant are as follows: The roots contain falcarinol, falcarindiol, panaxidol and polyacetylene 8-O-methylfalcarindiol.<sup>1,8</sup> The stem contains pectic polysaccharide (apiuman) containing d-galacturonic acid, 1-rhamnose, 1-arabinose and d-galactose.<sup>34</sup> Leaves contain 1-dodecanol, 9-octadecene-12-ynoic acid, methyl ester and tetradecene-1-ol acetate.<sup>31</sup> Celery seed contains caffeic acid, chlorogenic acid, apigenin, rutaretin, ocimene, bergapten and isopimpinellin.<sup>6</sup> The seed oil is composed of palmitic acid, stearic acid, oleic acid, linoleic acid, petroselinic acid, d-limonene, selinene, terpineol and santalol.<sup>33</sup>

Antioxidant Effect: In the study by Kolarovic et al,<sup>21</sup> the antioxidant activities[as measured by the content of reduced glutathione (GSH) and ferric reducing antioxidant power (FRAP)] of celery and parsley leaf and root juices in rats treated with doxorubicin, were investigated.

Percent (%)
57.7
18.7
8.6
8.1
2.4
0.5
0.3
0.3
0.3
0.2
0.1
0.1
0.1
0.1
0.1
0.1

Table 1	
Essential oils of A. graveolens L. seed,	GC/MS <sup>24</sup>

Celery root juice increased antioxidative capacity and the total antioxidative capacity (TAOC) in liver homogenate. Celery leaf juice increased GSH content but did not increase FRAP in liver homogenate. Study results show that celery increases antioxidant activity.

The study by Al Sa'aidi et al<sup>2</sup> of antioxidant activity of nbutanol celery extract (*Apium graveolens*) seed in streptozotocin-induced diabetic rats was investigated. Thirty-two mature male rats were divided into four groups as diabetic and non-diabetic. Rats  $\geq 200 \text{ mg/dl}$  of blood glucose were used as diabetic. Diabetic groups were drenched with drinking water, n-butanol extract (60 mg/kg, b.w.), or injected with insulin (4 IU/animal) respectively for 21 days. Blood and liver subcellular fluid were obtained for the evaluation of alanine aminotransferease (ALT), Aspartate aminotransferase (AST), catalase (CAT), Superoxide dismutase (SOD), Glutathione (GSH) transferase and -reductase enzymes and Malondialdehyde (MDA), glutathione concentrations.

N-butanol extract of celery seed or insulin therapy moderated blood glucose within a normal range, enhanced body weight gain and normalized the activities of all antioxidant enzymes. Study results show that n-butanol extract of celery seed has a potent role in ameliorating stressful complications accompanied by diabetes mellitus.

In the study by Li et al,<sup>29</sup> *in vitro* and *in vivo* antioxidant activity of ethanol extract of celery leaf was investigated. Superoxide dismutase (SOD), glutathione peroxidase (GSH-Px) and catalase and total antioxidant capacity (TAOC) activities were measured in serum, brain, heart, liver and kidneys. As a result, celery has a radical scavenging effect and SOD, GSH-Px CAT have been shown to significantly increase the activity.

Yıldız et al<sup>44</sup> identified the essential antioxidant compounds and measured the total antioxidant capacity with CUPRAC (cupric ion reducing antioxidant capacity) and ABTS spectrophotometric methods. The CUPRAC spectrophotometric method of TAC assay using copper(II)neocuproine (2,9-dimethyl-1,10- phenanthroline) was developed. Antioxidant compounds in celery plant extracted by HPLC were analyzed on one column of C18. Study results show that methanolic and ethanolic extract of celery leaves have antioxidant properties.

Yao et al<sup>43</sup> analyzed the phenolic compound composition and antioxidant activities of 11 celery varieties. The contents of total phenolics were measured using a Folin–Ciocalteu assay and the total antioxidant capacity was measured with the 1,1-diphenyl-2-picrylhydrazyl radical and 2,2-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) methods. The most common flavonoid in celery was apigenin and phenolic acid was p-coumaric acid.

The investigated celery varieties had high levels of phenolics and exhibited high antioxidant capacity. Antioxidant activity was found to be proportional to total flavonoids, total phenolic acids or total phenolics. In the study by Nagella et al<sup>31</sup> essential oil composition of celery leaf, immunotoxicity effects and antioxidant activity was investigated.

Essential oils contained in *A. graveolens* leaves were found using gas chromatography and mass spectroscopy (GC-MS). The essential oil from the *A. graveolens* leaves was investigated for scavenging of the 1,1-diphenyl-2picrylhydrazyl (DPPH) radical activity. The results showed that the essential oil from the *A. graveolens* has potential as a natural antioxidant and thus inhibits the unwanted oxidation process.

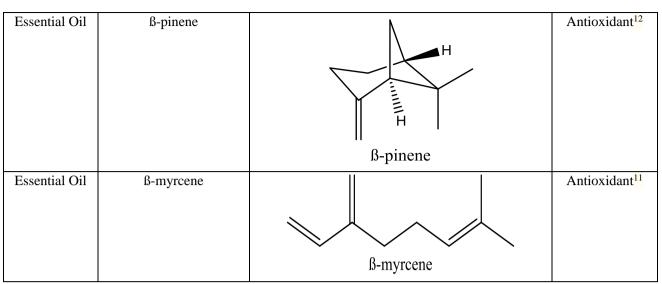
Shanmugapriya and Ushadevi<sup>38</sup> studied the antibacterial and antioxidant activity of Methanol, Diethyl ether and aqueous extracts of *Apium graveolens* seeds. The antioxidant activity of *A.graveolens* seed extracts was carried out 2, 2-Diphenyl-1- picrylhydrazyl (DPPH) assay method. The methanol extract showed the highest inhibition against bacterial pathogens and higher antioxidant activity than that of standard Gallic acid. Study result showed that *A. graveolens* seed extract is exhibiting enormous significance in therapeutic aspects. Uddin et al evaluated the phytochemical screenings, antioxidant activity and antimicrobial assay of *Apium graveolens* L.

The total phenolic content was slightly higher in methanolic fraction ( $63.46 \pm 12.00 \text{ mg GAE/g}$ ) than ethanol ( $36.60 \pm 12.28 \text{ mg GAE/g}$ ) and hexane fractions ( $34.86 \pm 6.96 \text{ mg GAE/g}$ ). The flavonoid content was high in methanolic extract ( $56.95 \pm 7.14 \text{ mg Quercetin/g}$ ). Ethanol extract showed good antimicrobial activity. Antioxidant activities of extracts were measured according to DPPH, ABTS and FRAP assays.

Group of Chemicals	Chemical Constituents	Structure	Reported Activity
Glycosides	Apigenin	НО	Antioxidant <sup>7,29</sup>
		OH opigenin	
Organic acid	Caffeic acid	HO HO caffeic acid	Antioxidant <sup>10</sup>
Organic acid ester	Chlorogenic acid	HO HO CATTER ACIA	Antioxidant <sup>36</sup>
Fatty acids	Palmitic acid	Chlorogenic acid	Antioxidant <sup>14,40</sup>
Essential Oil	Sedanolide	Palmitic acid	Antioxidant <sup>42</sup>
Essential Oil	Terpineol (2-(4- Methylcyclohex-3-en- 1-yl)propan-2-ol)	OH 2-(4-Methylcyclohex-3-en-1-yl)propan-2-ol	Antioxidant <sup>9</sup>

 Table 2

 The chemical constituent of the Apium graveolens L seed



Antioxidant activity assayed by FRAP was higher in methanolic fraction (12.48  $\pm$  1.06 mmole of FeSO4 equivalent/litre of extract) compared with other extracts. The study by Naglaa et al<sup>32</sup> for the constituents of the essential oil, antioxidant and antimicrobial activity of celery (*Apium graveolens*) was investigated. The chemical composition of the essential oils obtained by hydrodistillation was analyzed by GC/MS.

The antioxidant activities of volatile oils extracted from the celery were assessed by the Rancimat apparatus and DPPH. Study results show that all essential oils under study at various concentrations exhibited antioxidant activity.

Ksouda et al<sup>27</sup> investigated 25 Tunisian plant species of 13 families based on their oil and total phenolic contents. The phenolic content of seed methanolic extracts was measured by Folin–Ciocalteu assay (490  $\pm$  60 mg GAE/100 g Dry Weight). In the ABTS assay, the antioxidant activity value was 1000  $\pm$  150 mg TEAC/100 g DW. In the DPPH assay, the antioxidant activity value of *Apium graveolens* was 480  $\pm$  30 (mg TEAC/100 g DW).

The results showed that the seeds of *Apium graveolens* had high oil content, interesting fatty acid profiles and its methanolic extracts displayed high antioxidant capacities.<sup>19</sup> The leaves of *A. graveolens* were extracted with methanol and partitioned with water, ethyl acetate and butanol.

The phenolic content of the extracts was determined by Folin-Coicalteu method. Antioxidant capacity was measured by using  $\alpha$ ,  $\alpha$ -diphenyl- $\beta$ -picrylhydrazyl (DPPH),  $\beta$ -carotene-linoleate, reducing power, metal chelating effects and phosphomolybdenum method. The phenolic content of the extracts was expressed as gallic acid equivalents and was found to be highest in methanol (51.09 mg/g).

At concentration of 250 g/ml, methanol extract has the highest free radical scavenging activity and reducing power.

The study result showed that celery leaf vegetable is a good source of antioxidants due to its phenolic richness.

Han et al<sup>17</sup> investigated the effect of digestion on the phenolic compounds and antioxidant activity of celery leaf. 13 phenolic chemicals were discriminated by HPLC-MS and contents of phenolic and the antioxidant capacity were evaluated after digestion *in vitro*. The extraction of celery leaf decreased lipid peroxidation and reactive oxygen species level.

It was also found that celery leaf increased antioxidant activity of liver, spleen and thymus of mice treated with Dexamethasone. In the study by Popovic et al,<sup>35</sup> the potential protective action of the ether, chloroform, ethyl acetate, n-butanol and water extracts was assessed by the corresponding *in vitro* and *in vivo* tests.

In the *in vitro* experiments crude methanol extracts were tested as potential scavengers of free OH• and DPPH• radicals as well as inhibitors of liposomal peroxidation (LPx). The results showed that both the extracts of root and leaves are good scavengers of OH• and DPPH• radicals.

*In vivo* experiments were concerned with antioxidant systems (activities of GSHPx, GSHR, Px, CAT, SOD, GSH content and intensity of LPx) in liver homogenate and blood of mice after their treatment with extracts of celery leaves, or in combination with CCl<sub>4</sub>. On the basis of the results obtained, n-butanol extract showed the highest protective effect.

#### Conclusion

This study investigated the properties of celery leaves and seeds. Celery is a commercially important seed spice valued for its medicinal properties. Celery because of compounds such as coumarin, apigenin, luteolin, tannin, kaempferol has powerful antioxidant characteristics. The plant composition and medicinal properties need more research about its other useful and unknown properties.

Type of extract	Used Parts	Model	Results
Aqueous extract	Root and leaves <sup>21</sup>	In vivo	-Celery root juice increased antioxidative capacity, – Celery leaf juice increased GSH content
Aqueous extract	Seed <sup>2</sup>	In vivo	-n-Butanol extract of celery seed normalized the activities of all antioxidant enzymes
Ethanolic extract	Leaves <sup>29</sup>	In vivo and in vitro	-Scavenging activity on MDA and LPF. – Enhanced the activities of SOD, GSH- Px and CAT
Methanolic and Ethanolic extracts	Leaves <sup>44</sup>	In vitro	Increased total antioxidant capacity
Ethanolic extract	All of the parts <sup>43</sup>	In vitro	Excellent free radical scavenging activities
_	Leaves <sup>32</sup>	In vitro	Has potential as a natural antioxidant and thus inhibits unwanted oxidation process
Methanolic, diethyl ether and aqueous extracts	Seeds <sup>38</sup>	In vitro	Methanol extract showed the highest antioxidant activity
Methanolic, ethanol and hexane <sup>13</sup> extracts	_	In vitro	Antioxidant activity was observed
Aqueous extract	Seeds <sup>32</sup>	In vitro	Exhibited antioxidant activity
Methanolic extract	Seeds <sup>27</sup>	In vitro	Extract exhibited high antioxidant activity
Methanol, water, ethyl acetate and butanol extract	Leaves <sup>19</sup>	In vitro	The antioxidant and free radical scavenging activities of the extracts assayed through DPPH and reducing power were found to be highest with methanol
Water extract	Leaves <sup>17</sup>	In vitro and In vivo	The extraction of celery leaf decreased lipid peroxidation and reactive oxygen species level and elevated the antioxidant activities
Methanol, ethyl acetate, butanol and water extract	Root and leaves <sup>35</sup>	In vitro and In vivo	Root and leaves are good scavengers of OH• and DPPH• radicals and reduce liposomal peroxidation intensity in liposomes

 Table 3

 Summary of Antioxidant Activity of Celery

#### References

1. Al-Asmari A., Athar Md. T. and Kadasah S.G., An Updated Phytopharmacological Review on Medicinal Plant of Arab Region: *Apium graveolens* Linn., *Pharmacognosy Review*, doi:10.4103/ phrev.phrev\_35\_16, **11**, 13-18 (**2017**)

2. Al Sa'aidi J.A.A., Alrodhan M.N.A. and Ismael A.K., Antioxidant activity of n-butanol extract of celery (*Apium* graveolens) seed in streptozotocin-induced diabetic male rats, *Res Pharm Biotechnol.*, **4**, 24-29 (**2012**)

3. Asadi-Samani M., Kooti W., Aslani E. And Shirzad H., A systematic review of Iran's medicinal plants with anticancer effects, *J Evid Based Complementary Alternative Med.*, doi: 10. 1177/2156587215600873, **21**, 143-153 (**2016**)

4. Aydemir T. and Becerik S., Phenolic content and antioxidant activity of different extracts from *Ocimum Basilicum*, *Apium* 

*Graveolens* and *Lepidium Sativum* Seeds, *Journal of Food Biochemistry*, doi:10.1111/j.1745-4514.2010.00366.x, **35**, 62–79 (**2011**)

5. Barnes J., Anderson A.L. and Phillipson J.D., Herbal Medicines, Pharmaceutical Press: London, Chicago, 118 (**2002**)

6. Beier R., Ivie G., Oertli E. and Holt D., HPLC analysis of linear furocoumarins (psoralens) in healthy celery (*Apium graveolens*), *Food and Chemicak Toxicology*, doi:10.1016/0278-6915(83)90231-4, **21**, 163-165 (**1983**)

7. Birt D., Mitchell D., Gold B., Pour P. and Pinch H., Inhibition of ultraviolet light induced skin carcinogenesis in SKH-1 mice by apigenin, a plant flavonoid, *Anticancer Research*, **17**, 85–91 (**1996**)

8. Bohlmann F., Components of parsley and celery roots, *Chem. Ber.*, **100**, 3454-3456 (**1967**)

9. Burits M. and Bucar F., Antioxidant activity of Nigella sativa essential oil, *Phytother Res*, doi:10.1002/1099-1573(200008)14:5 <323::aid-ptr621> 3.0.co;2-q, **14**, 323–8 (**2000**)

10. Chen J.H. and Ho C.T., Antioxidant activities of caffeic acid and its related hydroxycinnamic acid compounds, *J Agric Food Chem.*, doi: 10. 1021/jf970055t, **45**, 2374–8 (**1997**)

11. Ciftci O., Ozdemir I., Tanyildizi S., Yildiz S. and Oguzturk H., Antioxidative effects of curcumin, ß-myrcene and 1,8-cineole against 2,3,7,8-tetrachlorodibenzo-p-dioxin-induced oxidative stress in rats liver, *Toxicol Ind Health*, doi: 10.1177/ 0748233710388452, **27**, 447–53 (**2011**)

12. Da Silva A.C., Lopes P.M., De Azevedo M.M., Costa D.C., Alviano C.S. and Alviano D.S., Biological activities of a-pinene and β-pinene enantiomers, *Molecules*, doi: 10.3390/molecules 17066305, **17**, 6305–16 (**2012**)

13. Din Z.U., Shad A.A., Bakht J., Ullah I. and Jan S., *In vitro* antimicrobial, antioxidant activity and phytochemical screening of *Apium graveolens*, *Pakistan Journal of Pharmaceutical Science*, **28**, 1699-1704 (**2015**)

14. Favre J. et al, Palmitic acid increases pro-oxidant adaptor protein p66Shc expression and affects vascularization factors in angiogenic mononuclear cells: Action of resveratrol, *Vascul Pharmacol.*, doi: 10.1016/j.vph.2015.08.003, **75**, 7–18 (**2015**)

15. Gauri M., Javed Ali S. and Shahid Khan M., A review of *Apium* graveolens (Karafs) with special reference to Unani medicine, *Int Arch Integr Med.*, **2**, 131-136 (**2015**)

16. Gutierrez R., Ramirez A., Campoy A., Flores J. and Flores S., Polyphenols of leaves of *Apium graveolens* inhibit in vitro protein glycation and protect RINm5F cells against methylglyoxalinduced cytotoxicity, *Functional Foods in Health and Disease*, doi: 10.31989/ffhd.v8i3.399, **8(3)**, 193-211 (**2018**)

17. Han L., Gao X., Xia T., Xueqian Z., Li X. and Gao W., Effect on digestion on the phenolic content and antioxidant activity of celery leaf and the antioxidant mechanism via Nrf2/HO-1 signaling pathways against Dexamethasone, *Journal of Food Biochemistry*, doi:10.1111/jfbc.12875, **43(7)** (**2019**)

18. Hazafa A., Rehman K., Jahan N. and Jabeen Z., The role of polyphenol (flavonoids) compounds in the treatment of cancer cells, *Nutrition and Cancer*, doi:10.1080/01635581.2019.163 7006, 1-12 (**2019**)

19. Jung W.S., Chung I.M., Kim S.H., Kim M.Y., Ahmad A. and Praveen N., In vitro antioxidant activity, total phenolics and flavonoids from celery (*Apium graveolens*) leaves, *Journal of Medicinal Plants Research*, **5(32)**, 7022-7030 (**2011**)

20. Khare C.P., Indian medicinal plants: London: Springer Science Pub. (2008)

21. Kolarovic J., Popovic M., Zlinská J., Trivic S. and Vojnovic M., Antioxidant activities of celery and parsley juices in rats treated with doxorubicin. Molecules, **15**, 6193-6204 (**2010**)

22. Kooti W., Akbari S., Samani M., Ghadery H. and Larky D., A review on medicinal plant of *Apium graveolens*, *Advanced Herbal Medicine*, **1**(1), 48-59 (**2014**)

23. Kooti W., Farokhipour M., Asadzadeh Z., Ashtary-Larky D. and Asadi-Samani M., The role of medicinal plants in the treatment of diabetes: a systematic review, *Electronic Physician*, doi: 10.19082/1832, **8**, 1832-1842 (**2016**)

24. Kooti W. et al, The effects of hydro-alcoholic extract of celery on lipid profile of rats fed a high fat diet, *Advances in Environmental Biology*, **8**, 325-330 (**2014**)

25. Kooti W., Moradi M., Ali-Akbari S., Sharafi-Ahvazi N., Asadi-Samani M. and Ashtary-Larky D., Therapeutic and pharmacological potential of *Foeniculum vulgare* Mill: a review, *Journal of Herb Med Pharmacology*, **4**, 1-9 (**2015**)

26 Kris-Etherton P.M., Hecker K.D., Bonanome A., Coval S.M., Binkoski A.E. and Hilpert K.F., Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer, *The American Journal of Medicine*, doi:10.1016/s0002-9343(01) 00995-0, **30**, 71-88 (**2002**)

27. Ksouda G., Hajji M., Sellimi S., Merlier F., Falcimaigne-Cordin A., Nasri M. and Thomasset B., A systematic comparison of 25 Tunisian plant species based on oil and phenolic contents, fatty acid composition and antioxidant activity, *Industrial Crops and Products*, doi: 10.1016/j.indcrop.2018.07.008, **123**, 768-778 (**2018**)

28. Labrador V., Freire P.F., Martin J.M.P. and Hazen M.J., Cytotoxicity of butylated hydroxyanisole in Vero cells, *Cell Biology and Toxicology*, doi: 10.1007/s10565-006-0153-6, **23**, 189-199 (**2007**)

29. Li P., Jia J., Zhang D., Xie J., Xu X. and Wei D., *In vitro* and *in vivo* antioxidant activities of a flavonoid isolated from celery (*Apium graveolens* L. var. dulce), *Food Funct.*, doi: 10.1039/c3fo60273g, **5**, 50-60 (**2014**)

30. Lone ZA, Lone Y, Khan S.S., Wani A.A. and Reshi M.I., Hepatoprotective medicinal plants used by the Gond and Bhill tribals of District Raisen Madhya Pradesh, India, *J Med Plants Res.*, doi:10.5897/jmpr2015.5764, **9**, 400-406 (**2015**)

31. Nagella P., Ahmad A., Kim S.J. and Chung I.M., Chemical composition, antioxidant activity and larvicidal effects of essential oil from leaves of *Apium graveolens*, *Immunopharmacol Immunotoxicol.*, doi: 10.3109/08923973.2011.592534, **34**, 205-209 (**2012**)

32. Naglaa H.M. et al, Antioxidant and antimicrobial activity of celery (*Apium graveolens*) and coriander (Coriandrum sativum) herb and seed essential oils, *International Journal of Current Microbiology and Applied Science*, **4**, 284-296 (**2015**)

33. Ngo-Duy C., Destaillats F., Keskitalo M., Arul J. and Angers P., Triacylglycerols of Apiaceae seed oils: Composition and regiodistribution of fatty acids, *European Journal of Lipid Science and Technology*, doi: 10.1002/ejlt.200800178, **111**, 164–169 (2009)

34. Petrova I., Petkova N., Kyobashieva K., Denev P., Simitchiev A., Todorova M. and Dencheva N., Isolation of Pectic Polysaccharides from Celery (*Apium graveolens var. rapaceum D.* C.) and Their Application in Food Emulsions, *Turkish Journal of Agricultural and Natural Sciences*, **2**, 1818-1824 (**2014**)

#### Res. J. Chem. Environ.

35. Popovic M., Kaurinovic B., Trivic S., Dukic N. and Bursac M., Effect of Celery (*Apium graveolens*) extracts on some biochemical parameters of oxidative stres in mice treated with carbon tetrachloride, *Phytotherapy Research*, doi: 10.1002/ptr.1871, **20**, 531–537 (**2006**)

36. Sato Y. et al, *In vitro* and *in vivo* antioxidant properties of chlorogenic acid and caffeic acid, *Int J Pharm.*, doi: 10.1016/j.ijpharm.2010.09.035, **403**, 136–8 (**2011**)

37. Sedigheh A. et al, Hypoglycaemic and hypolipidemic effects of pumpkin (*Cucurbita pepo L.*) on alloxan-induced diabetic rats, *Afr J Pharm Pharmacol.*, doi: 10.5897/AJPP11.635, **5(23)**, 2620-6 (**2011**)

38. Shanmugapriya R. and Ushadevi T., In vitro antibacterial and antioxidant activities of *Apium graveolens* L. seed extracts, *International Journal of Drug Development and Research*, **6**, 165-170 (**2014**)

39. Shirzad H., Taji F. and Rafieian-Kopaei M., Correlation between antioxidant activity of garlic extracts and WEHI-164 fibrosarcoma tumor growth in BALB/c mice, *J Med Food*, doi: 10.1089/jmf.2011.1594, **14(9)**, 969-74 (**2011**)

40. Shukla S., Mishra T., Pal M., Meena B., Rana T.S. and Upreti D.K., Comparative analysis of fatty acids and antioxidant activity of *Betula utilis* bark collected from different geographical region

of India, *Free Radic Antioxid.*, doi: 10.5530/fra.2017.1.12, **7**, 80-85 (**2017**)

41. Tauchen J., Huml L., Bortl L., Doskocil I., Jarosova V., Marsik P., Frankova A., Peralta Z., Zans M., Havlik J., Lapcik O. and Kokoska L., Screening of medicinal plants traditionally used in Peruvian Amazon for in vitro antioxidant and anticancer potential, *Natural Product Research*, doi: 10.1080/14786419.2018.1462180, **33**, 2718-2721 (**2019**)

42. Woods J.A., Jewell C. and O'Brien N.M., Sedanolide, a natural phthalide from celery seed oil: effect on hydrogen peroxide and tert-butyl hydroperoxide-induced toxicity in HepG2 and CaCo-2 human cell lines, *In Vitr Mol Toxicol.*, doi:10.1089/10979330 1753407984, **14**, 233–40 (**2001**)

43. Yao Y., Sang W., Zhou M. and Ren G., Phenolic composition and antioxidant activities of 11 celery cultivars, *Journal of Food Science*, doi:10.1111/j.1750-3841.2009.01392.x, **75**, C9-C13 (2010)

44. Yildiz L., Baskan K.S., Tütem E. and Apak R., Combined HPLC-CUPRAC (cupric ion reducing antioxidant capacity) assay of parsley, celery leaves and nettle, *Talanta*, doi:10.1016/j.talanta.2008.06.028, **77**, 304-313 (**2008**).

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