

Influence of arginase activity with some oxidative stress parameters in Iraqi vitiligo patients.

Alkinani Aymen Abdulsattar^{1*}, Zainulabdeen Jwan Abdulmohsin¹ and Salman Husam Ali²

1. Department of Chemistry, College of Sciences, University of Baghdad, Baghdad, IRAQ

2. Department of Dermatology and Venereology, College of Medicine, University of Baghdad, Baghdad, IRAQ

*aymenalkinani90@gmail.com

Abstract

Arginase enzyme has drawn more attention recently because this enzyme enters in several pathways of intermediary metabolism. The purpose of this study was to measure the activity of Arginase, total antioxidant capacity, total oxidant capacity and peroxynitrite level in serum of vitiligo patients. The activity of Arginase was determined and total antioxidant capacity and total oxidant capacity were also determined while peroxynitrite levels were also determined. These methods were applied to vitiligo patients and healthy individuals who matched in age and gender with patients.

A highly significant increase in both serum arginase activity and its specific activity levels $p=0.000$ in patients group was observed in comparison with their levels in corresponding healthy group. Total antioxidant capacity in sera of patients group was highly significant lower $p=0.000$ when compared to the healthy group, while highly significant increase in total oxidant capacity and peroxynitrite levels $p=0.000$ in vitiligo patients group were observed in comparison with their levels in corresponding healthy control group. Vitiligo disease affects indirectly on the activity of arginase due to imbalance between antioxidant and oxidant system which lead to increased activity and expression of arginase.

Keywords: Arginase, total oxidant capacity, total antioxidant capacity, peroxynitrite and vitiligo.

Introduction

Vitiligo is an acquired chronic skin disease characterized by loss and disorder in melanocytes functions from the epidermis²⁷. The distribution of this disease is from 0.5% to 1% of the general population⁵. The main etiology is still unclear, therefore some theories including autoinflammatory, autoimmune, neural, genetic, autocytotoxic, were proposed to explain vitiligo disease^{5,27}. One of most important theory is autocytotoxic theory, which emanates from clinical observations and laboratory investigations of melanocytes, skin biopsies, serum, and circulating peripheral mononuclear cells (PMNC) of vitiligo.

The oxidative stress theory of vitiligo pathogenesis is largely based on the notion that the entire epidermis of vitiligo accumulates H_2O_2 that may reach to one millimole

concentration⁵⁰⁻⁵². At this concentration, H_2O_2 is considered to be the primary intracellular reactive oxygen species (ROS) that leads to the death of the epidermal functional melanocytes. Multiple biochemical alterations in the epidermis and melanocytes of vitiligo have been built in principle on this notion of increased epidermal H_2O_2 concentration^{7,46,61}. Mitochondrial dysfunction seems to play a major role in vitiligo pathogenesis¹⁷, and its increased production of H_2O_2 in vitiligo skin is believed to be the culprit of H_2O_2 -induced mitochondrial DNA damage⁴² that eventually leads to melanocyte apoptosis/death⁵⁴.

The enzyme is crucially involved in various aspects of inflammation. Arginase has been shown to be either responsible for or to participate in, for example, inflammation-triggered immune dysfunction, tumour immune escape, fibrosis, immunosuppression and immunopathology of infectious diseases⁸.

Arginase (L-arginine amidinohydrolase, EC 3.5.3.1) catalyzes the reaction in which L-arginine is converted to L-ornithine and urea⁵⁸. Arginase enzyme is paid more attention because this enzyme enters in several pathways of intermediary metabolism. Arginase competes with inducible Nitric oxide synthase (iNOS; NOS2), the high output, the inducible pathway for increased production of NO, for L-Arginine, their common substrate in multiple cell types including endothelial cells. NO levels can fall, and this may be the result of enhanced Arginase activity¹⁴. In humans, two arginase isoenzymes have been identified, arginase 1 and arginase 2, that differ in cellular location and tissue distribution²³.

Arginase expression and activity can be regulated in many cell types including vascular endothelial cells, smooth muscle cells, and macrophages, by various cytokines⁶³. The presence of at least two distinct genes for arginase in mammals has been established where arginase AI is strongly expressed in the liver and AII is expressed in extrahepatic tissues^{18,21,36,56}. The hepatic arginase AI is a cytosolic enzyme that plays a fundamental role in the last step of the urea cycle and is cytokine-inducible in many cell types⁹. Arginase AI can limit substrate availability for high output NO synthesis in cells co-expressing iNOS^{6,34} and it is induced by hypoxia, lipopolysaccharide (LPS), and IL-13 in a variety of cells and tissues^{31,53}.

Extrahepatic arginase AII is a mitochondrial isoform, and its biological function has been the subject of considerable interest. It seems that this isoform provides a supply of ornithine, a crucial metabolite in the biosynthesis of

glutamic acid, proline and polyamines^{1,45,55} and also is also inducible by hypoxia, LPS, TNF-, IFN-, and 8-Bromo-cGMP^{10,47}. In inflammatory diseases, such as acute respiratory distress syndrome, NO production from L-Arginine via NOS is involved in the initial host response whereas L-ornithine production from L-Arginine via arginase is involved in healing³⁰. Since polyamines are vital for cell proliferation, it is possible that the increased level of ornithine, due to the elevated arginase activity, may be linked to the development of carcinogenesis⁴².

Our current study aimed to determine influence in the arginase activity levels prevalent among vitiligo patients and to discover the associations between the arginase activity and total antioxidant capacity, total oxidant capacity and peroxynitrite concentration in the Iraqi vitiligo patients

Material and Methods

Two main groups were included in this study; 65 patients diagnosed with vitiligo without treatment (P group) and 40 apparently healthy controls (C group) comparable for age and gender without a history of vitiligo were involved in the current study. Each group was subdivided according to gender, where P group was divided to 33 male patients group (PM group) and 32 female patients group (PF group) while C group was divided to 20 male group (CM group) and 20 female patients group (CF group).

The samples were collected from patients who were attending to Baghdad teaching hospital in Medical city, Iraq. Venous fasting blood samples were collected from an antecubital vein (8.00-10.00 a.m.). The obtained sample was centrifuged at 3000xg for 5min and the collected serum was used freshly to measure arginase activity, protein profile, total antioxidant capacity, total oxidant capacity and peroxynitrite concentration.

In the current study patients with pregnancy, diabetic, anaemia, lactation, history of cutaneous photosensitivity, eye cataract or skin, cancer, dermatitis, psoriasis, taking a potent antioxidant, history of alcohol intake, smokers were excluded. This study protocol was approved by the Ethics Committee of the College of Science/ University of Baghdad.

Serum arginase activity assay: The serum arginase activity was measured according to Zofia et al method⁴¹ which depends on the measurement of ornithine concentration as the end product of the enzymatic reaction. Levels of total protein and albumin in the serum of studied groups were measured by commercial kit from HUMAN Company. Globulin concentration in sera samples of the studied groups in this study was calculated from the following equation:

Globulin conc. (g/L) = Total protein conc. – Albumin conc.

Serum total antioxidant capacity and total oxidant capacity: Total oxidant status (TOS) and total antioxidant

status (TAS) in the serum of studied group were measured according to methods developed by Ereli^{19,20}.

Serum peroxynitrite concentration: Serum peroxynitrite concentration was determined in serum of vitiligo patients and healthy groups according to the Beckman et al cited by VanUffelen et al.⁵⁹

Results and Discussion

The mean \pm SD ages of the two groups (P and C) was 31.52 ± 7.24 and 33.1 ± 8.62 years respectively, there were non-significant differences ($p > 0.05$) in the mean values of total protein, albumin and globulin concentrations of all the two studied groups patients (P) and healthy control (C) as present in figure 1. A highly significant increase in both serum arginase activity and its specific activity levels $p=0.000$ in P group was observed in comparison with their levels in corresponding healthy C group as shown in figure 2.

Total antioxidant capacity in sera of P group was highly significantly lower $p=0.000$ when compared to the healthy C group (Figure 2) while highly significant increase in total oxidant capacity $p=0.000$ in vitiligo patients group (P group) was observed in comparison with their levels in corresponding healthy control group (C group) as shown in figure 2. The concentrations of peroxynitrite were measured in serum samples, the obtained results reflect the presence of highly significant increase in peroxynitrite concentration ($p=0.000$) when compared with patients group and the healthy group. These results are presented in figure 2.

Gender in present study showed that mean levels of serum arginase activity and arginase specific activity in PM showed the significant increase at $p=0.011$ and $p=0.016$ respectively and in PF group there was a significant increase at $p=0.006$ and $p=0.007$ respectively when compared with corresponding healthy group (CM and CF groups respectively). Meanwhile, the levels of total antioxidants capacity and total oxidant capacity in patients and healthy control groups were altered, the result showed a significant decrease in total antioxidants capacity in both PM and PF groups $p=0.015$ and $p=0.028$ respectively, while highly significant increase in total oxidants capacity in both MP and PF groups $p=0.000$ and $p=0.000$ respectively was observed when compared with corresponding healthy groups (CM and CF groups).

It is clear from table 1 when that peroxynitrite concentration between male and female of vitiligo patients with their corresponding healthy groups was observed when significant increase in peroxynitrite concentration is present in PM and PF groups (0.002 and $p=0.009$ respectively).

The results of comparison among serum of male with female of vitiligo patients show both arginase activity and arginase specific activity as non-significant increase $p=0.786$ and $p=0.79$ respectively and non-significant in peroxynitrite

concentration, total antioxidants capacity and total oxidant capacity $p=0.99$, $p=1$ and $p=0.994$ respectively.

The current study was in agreement with several authors for the TAC and TOC systems in vitiligo. They found that patients with vitiligo had significantly lower TAC and higher TOC than controls group^{2,4,33}. Other researchers illustrated that there was impairment in the antioxidant system of vitiligo which leads to low level of TAC in both serum and tissue.

Also patients with vitiligo have been shown to have various alterations in oxidant, and antioxidant parameters in both tissues and blood such as increase in H_2O_2 ², increase in lipid peroxidation products such as malondialdehyde¹⁶, low level of vitamins C and E¹⁶, increased or decreased^{3,16,28} activity of superoxide dismutase^{25,39}, low activity of catalase^{3,16} and glutathione peroxidase activity showed conflicted result: it may be normal⁴⁰, increased³⁹ or decreased^{16,36} activity. These alterations in results may be due to difference of samples (tissue and blood), type, and duration of this disease. Our skin also acts as a barrier between the environment and the body, it is represented as the first defense exposed to a different array of physical, chemical, and biological agents, many of which either are inborn oxidants or catalyze the generation of ROS²².

In the last decades, many studies suggested that hypersensitivity to OS is a crucial role in determining melanocyte degeneration^{2,16,24,29,32}.

Vitiligo is disordered or impaired melanocyte cells, the cause of this disease is not understood yet, and several hypotheses have been proposed to explain its pathogenesis including autoimmune, neural, autotoxic, self-destruction and inherent defect theories^{5,27}. So a single mechanism is not responsible for all cases of melanocyte damage in vitiligo, and OS is considered a possible pathogenic event in melanocyte loss which believed that the formation of ROS in the melanocytes destroys melanocyte and leads to this disease²⁹.

TAC and TOC are essential for establishing the health status of the body³⁷, therefore decreased antioxidant levels and increased oxidant levels may play essential roles in the damage of melanocytes observed in vitiligo patients⁴⁹. The above reason can explain the decreasing in TAC and increasing in TOC levels in sera of vitiligo patients indicated in the present study. that is because a shift in the equilibrium between antioxidant-oxidant levels which leads to impaired balance between these important parameters. Meanwhile the global antioxidant capacity of patients with vitiligo might have been exhausted by a defense mechanism against oxidant processes.

Results presented here demonstrate that arginase activity and its specific activity in the sera of vitiligo patients are markedly increased when compared with apparently healthy

individuals. Also, their levels were increased in both PM and PF patients group when compared with CM and CF respectively while no one mentioned the effect of arginase enzyme in the serum of the vitiligo disease. The increase in arginase activity in our study can be explained depending on two theories autotoxic and autoimmune as follows:

1. As mentioned above, the patients with vitiligo had significantly lower TAC and high TOC than control as revealed by our study and others studies^{2,4,24,33}.

The free radical such as ROS that generates from melanocytes is produced by its mitochondria which are regarded as the main source of free radicals and play a pivotal role in cell survival and apoptosis⁶². ROS are considered normal byproducts of aerobic metabolism, namely, superoxide anion, hydroxyl anion, NO and H_2O_2 . The superoxide anion is primarily metabolized by the mitochondrial Mn-SOD and the cytosolic Cu/Zn-SOD, generating H_2O_2 ⁴³.

The widely accepted current OS theory of vitiligo pathogenesis is largely based on the notion that the entire epidermis of vitiligo accumulates H_2O_2 that may reach one millimole concentration⁵⁰⁻⁵². At this concentration, H_2O_2 is considered to be the primary intracellular ROS that leads to the death of the epidermal functional melanocytes. Multiple biochemical alterations in the epidermis and melanocytes of vitiligo have been built in principle on this notion of increased epidermal H_2O_2 concentration^{46, 7, 61}. Mitochondrial dysfunction seems to play a major role in vitiligo pathogenesis¹⁷ and increased production of H_2O_2 in vitiligo skin is believed to be the culprit of H_2O_2 -induced mitochondrial DNA damage⁵⁴ that eventually leads to melanocyte apoptosis/death.

In animal, Thengchaisri et al⁵⁷ studied the effect of H_2O_2 on the arginase enzyme by treating coronary arterioles of porcine with H_2O_2 for 60 minutes. The results from his experiment were increased arginase I mRNA by 2-fold and increased its activity without altering eNOS expression. The increase in arginase enzyme may be due to rapidly conversion of H_2O_2 to hydroxyl radical shown to activate mammalian p38 mitogen-activated protein kinases (P38 MAPKs) by a wide range of cellular stresses as well as in response to inflammatory cytokines^{44,64} and also can activate cAMP⁶² pathways. Both P38 MAPKs and cAMP can cause arginase induction in some cells.^{13,60} It is speculated that these ROS-induced signaling cascades may be involved in the upregulation of arginase expression.

Therefore, H_2O_2 appears to upregulate the gene and protein expression of arginase I in the coronary arteriolar wall, especially in endothelial cells. At the present time, the mechanism underlying the upregulation of arginase remains unclear. Another free radical caused an increase in the activity of arginase is ONOO-. The unregulated activity of nitric oxide synthase produced more NO as published

recently¹⁵ and also increases in OS, TOC levels (which mean increase in ROS), It has been shown that H₂O₂ may lead to an increase in other ROS such as superoxide¹¹ which can react rapidly with any available NO to form ONOO- and this reaction occurs more rapidly than SOD converting superoxide (H₂O₂)³⁸ leading to an increased cellular redox stress²⁶.

Sankaralingam et al⁴⁸ reported the effect of arginase II expression and its activity by ONOO- in the vasculature of women with preeclampsia vasculature. In addition, endothelial cells incubated with plasma from women with preeclampsia show enhanced arginase II expression and activity as well as increased superoxide levels and evidence for increased ONOO- formation. They proved that:

1) FeTPPS is a ONOO- scavenger in the presence of the preeclamptic plasma to investigate if ONOO- generated by preeclamptic plasma was responsible for enhanced arginase

expression and its activity. The result was significant reduction in the arginase expression that was induced by preeclamptic plasma suggesting that ONOO- generated by preeclamptic plasma is responsible for the upregulation of arginase.

2) When exogenous ONOO- was used, arginase expression and activity were upregulated.

In addition, another study of Thengchaisri et al⁵⁷ showed increased arginase activity and arginase I expression in bovine aortic endothelial cells by oxidative species ONOO- and H₂O₂ through the pathway involving PKC activation of p115-Rho GEF and subsequent RhoA/Rho kinase¹² also was in agreement with Sankaralingam et al⁴⁸ and Thengchaisri et al⁵⁷. The dominance of tissue expression of arginase I or II may depend on organ, disease state, and species involved³⁵.

Table 1
Mean of arginase activity, arginase specific activity and Peroxynitrite level in serum samples according to gender.

Samples	Group	Arginase activity (U/L)	P value	Specific activity of arginase (U/g)	P value	Peroxynitrite (µmole/L)	P value	TAC	P value	TOC	P value
Male	CM (n=20)	8.93 ± 2.14	0.011	0.129 ± 0.031	0.016	4.43 ± 2.35	0.002	1.49 ± 0.29	0.015	9.94 ± 4.93	0.000
	PM (n=33)	15.4 ± 4.76		0.216 ± 0.067		8.61 ± 2.61		1.09 ± 0.31		14.12 ± 5.52	
Female	CF (n=20)	5.9 ± 1.55	0.006	0.085 ± 0.022	0.007	3.78 ± 0.89	0.009	1.51 ± 0.23	0.028	8.24 ± 3.88	0.000
	PF (n=32)	13.77 ± 6.94		0.194 ± 0.097		8.33 ± 3.28		1.08 ± 0.39		13.06 ± 4.79	
Vitiligo patients	PM (n=33)	15.4 ± 4.76	0.786	0.216 ± 0.067	0.79	8.61 ± 2.61	0.99	1.09 ± 0.31	1	14.12 ± 5.52	0.994
	PF (n=32)	13.77 ± 6.94		0.194 ± 0.097		8.33 ± 3.28		1.08 ± 0.39		13.06 ± 4.79	

TAC (total antioxidant capacity), TOC (total oxidant capacity), CM (male healthy group), CF (female healthy group), PM (male patients group), PF (female patients group), PT1 (before treatment patients), PT2 (after treatment patients)
 p<0.05 significant, p<0.001 highly significant, p>0.05 non-significant

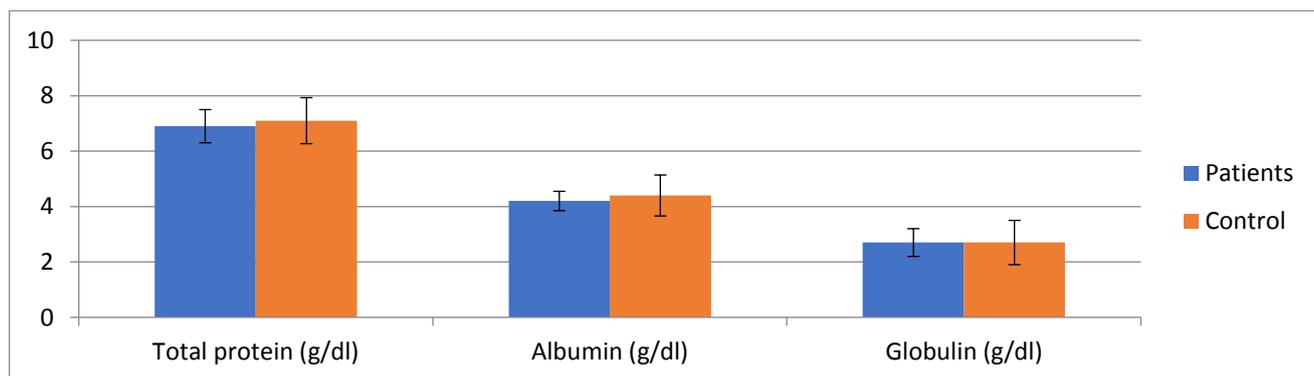


Figure 1: Mean of total protein, albumin and globulin concentrations of the two studied groups.

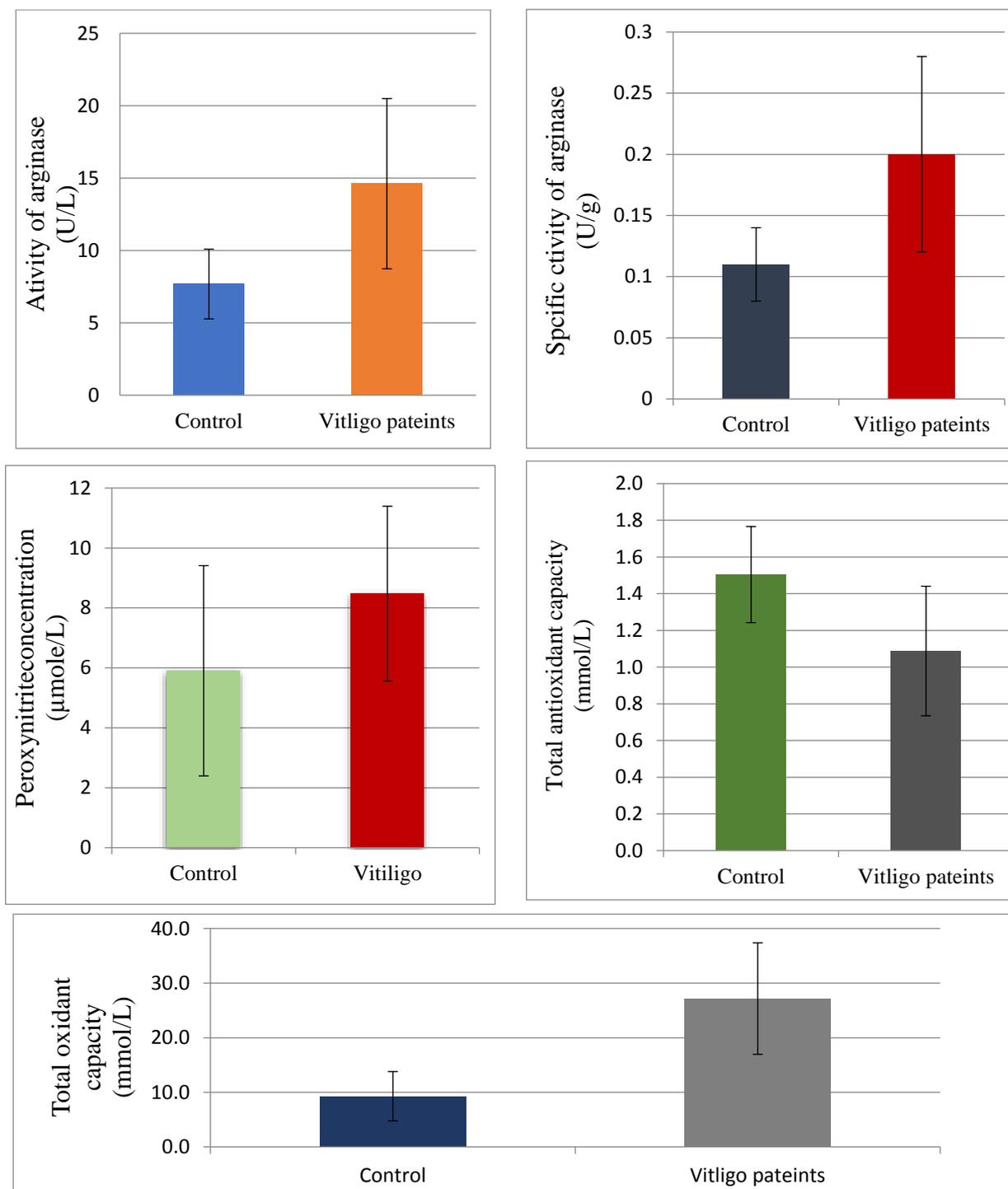


Figure 2: Comparison of serum arginase activity, arginase specific activity, peroxynitrite concentration, total antioxidants capacity and total oxidant capacity in sera of the vitiligo patients and control groups.

Conclusion

We measured arginase activity in the vitiligo and healthy individuals. Out of the result from the present study, one can conclude that vitiligo disease caused alterations in the activity of arginase activity and imbalance between antioxidant and oxidant system leading to increase in arginase activity and expression of arginase. Increased arginase activity led to increase both oxidative stress and inflammation and results in more destruction or dysregulation in melanocyte cells.

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References

1. Abraham A.K. and Pihl A., Role of polyamines in macromolecular synthesis, *Trends Biochem. Sci.*, **6**, 106-107 (1981)
2. Akoglu G., Emre S., Metin A., Akbas A., Yorulmaz A., Isikoglu S., Sener S. and Kilinc F., Evaluation of total oxidant and

antioxidant status in localized and generalized vitiligo, *Clin. Exp. Dermatol.*, **38(7)**, 701-706 (2013)

3. Arican O. and Kurutas E.B., Oxidative stress in the blood of patients with active localized vitiligo, *ACTA Dermatovenerologica Alpina Panonica et Adriatica*, **17(1)**, 12 (2008)

4. Boisseau-Garsaud A.M., Garsaud P., Lejoly-Boisseau H., Robert M., Quist D. and Arveiler B., Increase in total blood antioxidant status and selenium levels in black patients with active vitiligo, *Int. J. Dermatol.*, **41(10)**, 640-642 (2002)

5. Boniface K., Jacquemin C., Darrigade A.S., Dessarthe B., Martins C., Boukhedoui N., Vernisse C., Grasseau A., Thiolat D. and Rambert J., Vitiligo Skin Is Imprinted with Resident Memory CD8 T Cells Expressing CXCR3, *J. Invest. Dermatol.*, **138(2)**, 355-364 (2018)

6. Boucher J., Moali C. and Tenu J., Nitric oxide biosynthesis, nitric oxide synthase inhibitors and arginase competition for L-arginine utilization, *Cellular and Molecular Life Sciences CMLS*, **55(8-9)**, 1015-1028 (1999)

7. Briganti S., Caron-Schreinemachers A.L., Picardo M. and Westerhof W., Anti-oxidant defence mechanism in vitiliginous skin increases with skin type, *J. Eur. Acad. Dermatol. Venereol.*, **26(10)**, 1212-1219 (2012)

8. Bronte V. and Zanollo P., Regulation of immune responses by L-arginine metabolism, *Nat. Rev. Immunol.*, **5(8)**, 641-654 (2005)

9. Bruch-Gerharz D., Schnorr O., Suschek C., Beck K.F., Pfeilschifter J., Ruzicka T. and Kolb-Bachofen V., Arginase 1 Overexpression in Psoriasis: Limitation of Inducible Nitric Oxide Synthase Activity as a Molecular Mechanism for Keratinocyte Hyperproliferation, *The American Journal of Pathology*, **162(1)**, 203-211 (2003)

10. Buga G.M., Singh R., Pervin S., Rogers N.E., Schmitz D.A., Jenkinson C.P., Cederbaum S.D. and Ignarro L.J., Arginase activity in endothelial cells: inhibition by NG-hydroxy-L-arginine during high-output NO production, *Am. J. Physiol.*, **271(5 Pt 2)**, H1988-1998 (1996)

11. Carter W., Narayanan P.K. and Robinson J., Intracellular hydrogen peroxide and superoxide anion detection in endothelial cells, *J. Leukoc. Biol.*, **55(2)**, 253-258 (1994)

12. Chandra S., Romero M., Shatanawi A., Alkilany A., Caldwell R.B. and Caldwell R.W., Oxidative species increase arginase activity in endothelial cells through the RhoA/Rho kinase pathway, *Br. J. Pharmacol.*, **165(2)**, 506-519 (2012)

13. Chang C.I., Zoghi B., Liao J.C. and Kuo L., The involvement of tyrosine kinases, cyclic AMP/protein kinase A and p38 mitogen-activated protein kinase in IL-13-mediated arginase I induction in macrophages: its implications in IL-13-inhibited nitric oxide production, *The Journal of Immunology*, **165(4)**, 2134-2141 (2000)

14. Chang C.I., Liao J.C. and Kuo L., Arginase modulates nitric oxide production in activated macrophages, *Am. J. Physiol.*, **274(1 Pt 2)**, H342-348 (1998)

15. Cortas N.K. and Wakid N.W., Determination of inorganic nitrate in serum and urine by a kinetic cadmium-reduction method, *Clin. Chem.*, **36(8)**, 1440-1443 (1990)

16. Dammak I., Boudaya S., Ben Abdallah F., Turki H., Attia H. and Hentati B., Antioxidant enzymes and lipid peroxidation at the tissue level in patients with stable and active vitiligo, *Int. J. Dermatol.*, **48(5)**, 476-480 (2009)

17. Dell'Anna M.L., Urbanelli S., Mastrofrancesco A., Camera E., Iacovelli P., Leone G., Manini P., D'ischia M. and Picardo M., Alterations of mitochondria in peripheral blood mononuclear cells of vitiligo patients, *Pigment Cell Res.*, **16(5)**, 553-559 (2003)

18. Dizikes G.J., Grody W.W., Kern R.M. and Cederbaum S.D., Isolation of human liver arginase cDNA and demonstration of nonhomology between the two human arginase genes, *Biochem. Biophys. Res. Commun.*, **141(1)**, 53-59 (1986)

19. Erel O., A novel automated method to measure total antioxidant response against potent free radical reactions, *Clin. Biochem.*, **37(2)**, 112-119 (2004)

20. Erel O., A new automated colorimetric method for measuring total oxidant status, *Clin. Biochem.*, **38(12)**, 1103-1111 (2005)

21. Gotoh T., Araki M. and Mori M., Chromosomal localization of the human arginase II gene and tissue distribution of its mRNA, *Biochem. Biophys. Res. Commun.*, **233(2)**, 487-491 (1997)

22. GÜNTAŞ G., Engin B., Ekmekçi Ö.B., Kutlubay Z., Ekmekci H., Songür A., Uzunçakmak T.K.Ü., Vehid H.E., Serdaroğlu S. and Tüzün Y., Evaluation of advanced oxidation protein products, prooxidant-antioxidant balance and total antioxidant capacity in untreated vitiligo patients, *Annals of Dermatology*, **27(2)**, 178-183 (2015)

23. Guoyao W. and Morris S.M., Arginine metabolism: nitric oxide and beyond, *Biochem. J.*, **336(1)**, 1-17 (1998)

24. Hofny E.R., Hasan H.I., Mohamed N.A., Bamatraf M.S. and Mahran A.M., Serum and tissue levels of total antioxidant capacity in nonsegmental vitiligo, *Journal of the Egyptian Women's Dermatologic Society*, **15(1)**, 35-39 (2018)

25. Jain A., Mal J., Mehndiratta V., Chander R. and Patra S.K., Study of oxidative stress in vitiligo, *Indian J. Clin. Biochem.*, **26(1)**, 78-81 (2011)

26. Katusic Z.S., Vascular endothelial dysfunction: does tetrahydrobiopterin play a role?, *American Journal of Physiology-Heart and Circulatory Physiology*, **281(3)**, H981-H986 (2001)

27. Khurram H. and AlGhamdi K.M., The relationship between the serum level of vitamin D and vitiligo: a controlled study on 300 subjects, *J. Cutan. Med. Surg.*, **20(2)**, 139-145 (2016)

28. Koca R., Armutcu F., Altinyazar H. and Gürel A., Oxidant-antioxidant enzymes and lipid peroxidation in generalized vitiligo, *Clinical and Experimental Dermatology: Experimental Dermatology*, **29(4)**, 406-409 (2004)

29. Laddha N.C., Dwivedi M., Mansuri M.S., Gani A.R., Ansarullah M., Ramachandran A., Dalai S. and Begum R., Vitiligo: interplay between oxidative stress and immune system, *Exp. Dermatol.*, **22(4)**, 245-250 (2013)

30. Li H., Meininger C.J., Kelly K.A., Hawker J.R., Morris S.M. and Wu G., Activities of arginase I and II are limiting for

- endothelial cell proliferation, *Am. J. Physiol. Regul. Integr. Comp. Physiol.*, **282**(1), R64-69 (2002)
31. Louis C.A., Reichner J.S., Henry W.L., Mastrofrancesco B., Gotoh T., Mori M. and Albina J.E., Distinct arginase isoforms expressed in primary and transformed macrophages: regulation by oxygen tension, *Am. J. Physiol.*, **274**(3 Pt 2), R775-782 (1998)
32. Maresca V., Roccella M., Roccella F., Camera E., Del Porto G., Passi S., Grammatico P. and Picardo M., Increased sensitivity to peroxidative agents as a possible pathogenic factor of melanocyte damage in vitiligo, *J. Invest. Dermatol.*, **109**(3), 310-313 (1997)
33. Mehaney D.A., Darwish H.A., Hegazy R.A., Nooh M.M., Tawdy A.M., Gawdat H.I. and El-Sawalhi M.M., Analysis of oxidative stress status, catalase and catechol-O-methyltransferase polymorphisms in Egyptian vitiligo patients, *PLoS One*, **9**(6), e99286 (2014)
34. Mori M. and Gotoh T., Relationship between arginase activity and nitric oxide production, *Nitric Oxide*, <https://doi.org/10.1016/B978-012370420-7/50013-7>, 199-208 (2000)
35. Morris S.M., Recent advances in arginine metabolism: roles and regulation of the arginases, *Br. J. Pharmacol.*, **157**(6), 922-930 (2009)
36. Ohtake A., Takiguchi M., Shigeto Y., Amaya Y., Kawamoto S. and Mori M., Structural organization of the gene for rat liver-type arginase, *J. Biol. Chem.*, **263**(5), 2245-2249 (1988)
37. Ozyazgan S., Andican G., Erman H., Tuzcu A., Uzun H., Onal B. and Ozyazgan Y., Relation of protein oxidation parameters and disease activity in patients with Behçet's disease, *Clinical Laboratory*, **59**(7-8), 819-825 (2013)
38. Pacher P., Beckman J.S. and Liaudet L., Nitric oxide and peroxynitrite in health and disease, *Physiol. Rev.*, **87**(1), 315-424 (2007)
39. Passi S., Grandinetti M., Maggio F., Stancato A. and De Luca C., Epidermal oxidative stress in vitiligo, *Pigment Cell Res.*, **11**(2), 81-85 (1998)
40. Picardo M., Passi S., Morrone A., Grandinetti M., Carlo A.D. and Ippolito F., Antioxidant status in the blood of patients with active vitiligo, *Pigment Cell Res.*, **7**(2), 110-115 (1994)
41. Poremska Z. and Kedra M., Early diagnosis of myocardial infarction by arginase activity determination, *Clin. Chim. Acta*, **60**(3), 355-361 (1975)
42. Poremska Z., Luboiński G., Chrzanowska A., Mielczarek M., Magnuska J. and Barańczyk-Kuźma A., Arginase in patients with breast cancer, *Clin. Chim. Acta*, **328**(1-2), 105-111 (2003)
43. Raha S., McEachern G.E., Myint A.T. and Robinson B.H., Superoxides from mitochondrial complex III: the role of manganese superoxide dismutase, *Free Radical Biol. Med.*, **29**(2), 170-180 (2000)
44. Ramesh G. and Reeves W.B., p38 MAP kinase inhibition ameliorates cisplatin nephrotoxicity in mice, *American Journal of Physiology-Renal Physiology*, **289**(1), F166-F174 (2005)
45. Russell D.H. and McVicker T.A., Polyamine biogenesis in the rat mammary gland during pregnancy and lactation, *Biochem. J.*, **130**(1), 71-76 (1972)
46. Salem M.M., Shalhaf M., Gibbons N.C., Chavan B., Thornton J. and Schallreuter K.U., Enhanced DNA binding capacity on up-regulated epidermal wild-type p53 in vitiligo by H₂O₂-mediated oxidation: a possible repair mechanism for DNA damage, *The FASEB Journal*, **23**(11), 3790-3807 (2009)
47. Nagasaki Salimuddin A., Gotoh T., Isobe H. and Mori M., Regulation of the genes for arginase isoforms and related enzymes in mouse macrophages by lipopolysaccharide, *Am. J. Physiol.*, **277**(1 Pt 1), E110-117 (1999)
48. Sankaralingam S., Xu H. and Davidge S.T., Arginase contributes to endothelial cell oxidative stress in response to plasma from women with preeclampsia, *Cardiovasc. Res.*, **85**(1), 194-203 (2009)
49. Schallreuter K., Successful treatment of oxidative stress in vitiligo, *Skin Pharmacol. Physiol.*, **12**(3), 132-138 (1999)
50. Schallreuter K.U., Moore J., Behrens-Williams S., Panske A. and Harari M., Rapid initiation of repigmentation in vitiligo with Dead Sea climatotherapy in combination with pseudocatalase (PC-KUS), *Int. J. Dermatol.*, **41**(8), 482-487 (2002)
51. Schallreuter K.U., Moore J., Wood J.M., Beazley W.D., Gaze D.C., Tobin D.J., Marshall H.S., Panske A., Panzig E. and Hibberts N.A., In vivo and in vitro evidence for hydrogen peroxide (H₂O₂) accumulation in the epidermis of patients with vitiligo and its successful removal by a UVB-activated pseudocatalase, *Journal of Investigative Dermatology Symposium Proceedings*, Elsevier (1999)
52. Schallreuter K.U., Moore J., Wood J.M., Beazley W.D., Peters E.M., Marles L.K., Behrens-Williams S.C., Dummer R., Blau N. and Tho Ény B., Epidermal H₂O₂ accumulation alters tetrahydrobiopterin (6BH4) recycling in vitiligo: identification of a general mechanism in regulation of all 6BH4-dependent processes?, *J. Invest. Dermatol.*, **116**(1), 167-174 (2001)
53. Sonoki T., Nagasaki A., Gotoh T., Takiguchi M., Takeya M., Matsuzaki H. and Mori M., Coinduction of nitric-oxide synthase and arginase I in cultured rat peritoneal macrophages and rat tissues in vivo by lipopolysaccharide, *J. Biol. Chem.*, **272**(6), 3689-3693 (1997)
54. Swalwell H., Latimer J., Haywood R.M. and Birch-Machin M.A., Investigating the role of melanin in UVA/UVB- and hydrogen peroxide-induced cellular and mitochondrial ROS production and mitochondrial DNA damage in human melanoma cells, *Free Radical Biol. Med.*, **52**(3), 626-634 (2012)
55. Tabor C.W. and Tabor H., Polyamines in microorganisms, *Microbiol. Rev.*, **49**(1), 81 (1985)
56. Takiguchi M., Haraguchi Y. and Mori M., Human liver-type arginase gene: structure of the gene and analysis of the promoter region, *Nucleic Acids Res.*, **16**(18), 8789-8802 (1988)
57. Thengchaisri N., Hein T.W., Wang W., Xu X., Li Z., Fossum T.W. and Kuo L., Upregulation of arginase by H₂O₂ impairs

endothelium-dependent nitric oxide-mediated dilation of coronary arterioles, *Arterioscler. Thromb. Vasc. Biol.*, **26(9)**, 2035-2042 (2006)

58. van den Berg M.P., Meurs H. and Gosens R., Targeting arginase and nitric oxide metabolism in chronic airway diseases and their co-morbidities, *Curr. Opin. Pharmacol.*, **40**, 126-133 (2018)

59. Van Uffelen E.B., de Koster M. B., Van Steveninck J. and Elferink G.J., Intracellular but not extracellular conversion of nitroxyl anion into nitric oxide leads to stimulation of human neutrophil migration, *Biochem. J.*, **330(2)**, 719-722 (1998)

60. Wei L.H., Jacobs A.T., Morris Jr. S.M. and Ignarro L.J., IL-4 and IL-13 upregulate arginase I expression by cAMP and JAK/STAT6 pathways in vascular smooth muscle cells, *American Journal of Physiology-Cell Physiology*, **279(1)**, C248-C256 (2000)

61. Yao L., Hu D.N., Chen M. and Li S.S., Subtoxic levels hydrogen peroxide-induced expression of interleukin-6 by epidermal melanocytes, *Archives of Dermatological Research*, **304(10)**, 831-838 (2012)

62. Zailaie M.Z., Epidermal hydrogen peroxide is not increased in lesional and non-lesional skin of vitiligo, *Archives of Dermatological Research*, **309(1)**, 31-42 (2017)

63. Zhang C., Hein T.W., Wang W., Chang C.I. and Kuo L., Constitutive expression of arginase in microvascular endothelial cells counteracts nitric oxide-mediated vasodilatory function, *FASEB J.*, **15(7)**, 1264-1266 (2001)

64. Zhang Z., Leonard S.S., Huang C., Vallyathan V., Castranova V. and Shi X., Role of reactive oxygen species and MAPKs in vanadate-induced G2/M phase arrest, *Free Radical Biol. Med.*, **34(10)**, 1333-1342 (2003).